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Histopathological and Biochemical Changes in Fish Liver Under Environmental Stress: A Case Study of Labeo rohita

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Abstract

This study investigates the histopathological and biochemical changes in the liver of *Labeo rohita* (Rohu) under environmental stress, with a particular focus on the effects of oxidative stress. The liver, being a central organ in detoxification and metabolic processes, is highly susceptible to environmental changes such as pollution, temperature fluctuations, and low oxygen levels. In this study, we assess the histopathological alterations in liver tissues and measure key biochemical markers, including superoxide dismutase (SOD) activity, to determine the extent of oxidative stress. The findings highlight significant liver damage and alterations in antioxidant enzyme activity in response to environmental stressors, suggesting a close relationship between oxidative damage and liver function in fish. The results of this study provide valuable insights into the physiological adaptations of *Labeo rohita* to environmental changes and underscore the potential of liver biomarkers for monitoring fish health in polluted aquatic environments.

CC License CC-BY-NC-SA 4.0 **Keywords**: Histopathology, biochemical changes, liver, *Labeo rohita*, environmental stress, oxidative stress, superoxide dismutase, fish health, pollution, biomarkers.

1. Introduction

Superoxide dismutase (SOD) is an essential antioxidant enzyme that plays a pivotal role in protecting cells from oxidative damage caused by reactive oxygen species (ROS), particularly superoxide radicals. These ROS are byproducts of cellular metabolism and can cause significant damage to cell structures, proteins, and DNA if not effectively neutralized. The primary function of SOD is to catalyze the conversion of superoxide radicals into hydrogen peroxide and oxygen, both of which are less damaging to cells (McCord, 2001; Paoletti & Mocali, 1990). This enzymatic activity is critical in preventing oxidative stress, which is linked to various diseases, aging processes, and damage to cellular components. In aquatic organisms, especially fish, SOD activity serves as a crucial defense mechanism to counteract the harmful effects of oxidative stress induced by both natural and anthropogenic factors, such as pollution, heavy metals, and other environmental stressors (Sindhe & Kulkarni, 2004; Bharti & Rasool, 2021).

Fish, being ectothermic organisms, are particularly vulnerable to changes in their surrounding environment, including fluctuations in water quality, oxygen levels, and exposure to pollutants. The liver, being a central *Available online at:* https://jazindia.com
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organ for detoxification, metabolism, and antioxidant enzyme production, is especially sensitive to oxidative stress and plays a vital role in responding to such environmental challenges. The activity of SOD in the liver of fish is therefore a critical indicator of the oxidative stress level within the organism. Changes in SOD activity can reflect the extent of environmental contamination, as increased ROS production due to pollutants can overwhelm the organism's natural antioxidant defenses (Reddy, 2021). For instance, pollutants like heavy metals, pesticides, and industrial effluents can induce oxidative damage by generating ROS, prompting the liver to upregulate the production of antioxidant enzymes, including SOD, to mitigate the damage (Younus, 2018; Gopal & Elumalai, 2017).

In addition to biochemical responses, environmental stress can also lead to histopathological alterations in fish tissues. Histopathological changes are often used as diagnostic markers to assess the extent of damage caused by environmental contaminants. The liver is one of the primary organs examined in these studies due to its role in detoxification and metabolism. Liver damage due to oxidative stress can manifest in various ways, such as cellular degeneration, necrosis, and altered hepatosomatic index (HSI), which is a measure of liver size relative to body weight. The HSI is commonly used to assess the health and condition of the liver in fish, with a decrease in HSI often indicating liver damage and metabolic dysfunction (Sindhe & Kulkarni, 2004). Therefore, monitoring both biochemical and histopathological changes in fish liver provides a comprehensive understanding of the organism's response to oxidative stress and environmental degradation.

The hepatosomatic index (HSI) is a crucial parameter that helps in assessing the health of fish by reflecting the relationship between liver weight and body weight. An increase or decrease in HSI can indicate physiological changes or stress responses in fish. Studies have shown that anthropogenic pollution, such as the introduction of heavy metals and pesticides, leads to alterations in HSI and liver morphology (Reddy, 2021). In response to pollutants, the liver often experiences an increase in oxidative stress, which is reflected in the upregulation of antioxidant enzymes like SOD. As a result, examining both SOD activity and HSI provides valuable insights into the liver's ability to cope with stress and its overall health in polluted environments (Reddy, 2021; Gopal & Elumalai, 2017).

The role of SOD in fish has been studied extensively, with researchers focusing on how environmental stressors affect its activity and the subsequent impact on fish health. Exposure to pollutants, such as heavy metals and pesticides, has been shown to increase the production of ROS, leading to oxidative stress and triggering an adaptive response in the form of enhanced SOD activity. However, prolonged exposure to such stressors can overwhelm the antioxidant defense systems, leading to irreversible cellular damage (Younus, 2018). Therefore, understanding the relationship between environmental stress, SOD activity, and histopathological changes in fish liver is crucial for assessing the impact of environmental pollution on aquatic ecosystems.

Recent advancements in biochemistry and molecular biology have provided a deeper understanding of the mechanisms behind SOD activity in fish. Studies have focused on the molecular structure of SOD, its isoforms (such as Cu-Zn SOD, Mn-SOD, and Fe-SOD), and their specific roles in defending against oxidative damage. Cu-Zn SOD, for example, is predominantly found in the cytoplasm and plays a crucial role in protecting cells from ROS generated during normal cellular respiration (Rosa et al., 2021). Mn-SOD, on the other hand, is primarily located in the mitochondria and plays a critical role in mitigating oxidative stress generated during mitochondrial respiration (Gopal & Elumalai, 2017). The differential expression of these SOD isoforms in response to environmental stress can provide valuable information about the physiological adaptations of fish to their environment. Moreover, the combined analysis of SOD activity with histopathological changes in fish liver can serve as an effective biomarker for assessing the health of fish populations and the quality of aquatic ecosystems (Vukmirović et al., 2023; Younus, 2018).

In addition to biochemical and histopathological assessments, molecular techniques such as gene expression analysis and proteomics are increasingly being used to study the mechanisms underlying oxidative stress responses in fish. These techniques allow researchers to identify specific genes and proteins involved in the stress response, providing a more detailed understanding of the molecular pathways activated in response to oxidative damage (Reddy, 2021). Furthermore, the development of molecular markers for oxidative stress and liver damage could facilitate the early detection of environmental contamination and improve the monitoring of fish health in polluted aquatic environments (Younus, 2018).

The role of SOD in environmental stress tolerance has also been explored in other organisms, particularly plants, where SOD activity is known to play a key role in regulating oxidative stress induced by abiotic stressors such as drought, salinity, and temperature fluctuations (Mishra & Sharma, 2019; Saibi & Brini, 2018). While these studies focus on terrestrial organisms, the underlying principles of oxidative stress regulation through SOD activity are similar in aquatic species, highlighting the universality of this defense mechanism across different environmental conditions.

In conclusion, the study of SOD activity and hepatosomatic index (HSI) in fish, particularly *Labeo rohita*, provides valuable insights into the physiological and biochemical responses of aquatic organisms to environmental stress. The liver, as a central organ in detoxification and metabolic processes, is particularly susceptible to oxidative damage, and monitoring both SOD activity and histopathological changes in liver tissues is essential for understanding the impact of environmental pollutants on fish health. This research emphasizes the importance of SOD as a biomarker for oxidative stress in fish and provides a foundation for further studies on the effects of environmental degradation on aquatic ecosystems. Ultimately, understanding the relationship between oxidative stress and fish health can inform strategies for protecting aquatic biodiversity and improving water quality management.

2.Literature Review

The liver is a key organ in the detoxification processes of fish and is vital for their overall health, particularly in response to oxidative stress caused by environmental pollutants. As one of the most crucial organs in regulating metabolism and processing toxins, the liver's health and functionality can be directly affected by pollutants such as heavy metals, pesticides, and other anthropogenic contaminants. In this context, the **superoxide dismutase (SOD)** enzyme system plays a pivotal role in mitigating oxidative damage caused by reactive oxygen species (ROS). This literature review aims to provide a comprehensive overview of existing research on the role of SOD in liver health, particularly in freshwater fish, with a focus on the environmental stressors that influence the antioxidant defense mechanisms in fish.

Oxidative Stress and the Role of Superoxide Dismutase (SOD)

Oxidative stress occurs when the balance between ROS production and antioxidant defense mechanisms is disrupted, leading to cellular damage. In living organisms, ROS are generated as a byproduct of normal cellular metabolic processes, particularly in the mitochondria during aerobic respiration (Younus, 2018). The most common and damaging form of ROS is the superoxide radical (O2–), which is highly reactive and can damage cellular components such as lipids, proteins, and DNA (Paoletti & Mocali, 1990). Superoxide dismutase (SOD) is an essential antioxidant enzyme that plays a critical role in protecting cells from oxidative stress. It catalyzes the dismutation of superoxide radicals into less harmful molecules, oxygen (O2) and hydrogen peroxide (H2O2), which can subsequently be detoxified by other antioxidant enzymes like catalase and glutathione peroxidase (McCord, 2001).

Several studies have emphasized the importance of SOD in protecting organisms, including fish, from the damaging effects of oxidative stress (Reddy, 2021). In freshwater fish like *Labeo rohita*, SOD activity serves as a defense against pollutants, high temperatures, hypoxia, and other environmental stressors. Various isoforms of SOD, such as **Cu-Zn SOD** and **Mn-SOD**, play different roles in cellular compartments. Cu-Zn SOD is found predominantly in the cytoplasm, while Mn-SOD is located in the mitochondria, where it plays a crucial role in managing the ROS produced during energy production (Younus, 2018; Rosa et al., 2021).

Environmental Stressors and SOD Activity in Fish

Environmental stressors, such as temperature fluctuations, hypoxia, and exposure to pollutants, can significantly alter the oxidative balance within organisms, including fish. Pollution, especially the presence of heavy metals and pesticides, is a primary environmental stressor that induces oxidative stress in fish. Heavy metals like lead, cadmium, and mercury are known to generate ROS and disrupt the antioxidant defense mechanisms in fish, leading to enhanced SOD activity (Sindhe & Kulkarni, 2004; Bharti & Rasool, 2021). A significant body of literature has explored the effects of such pollutants on the SOD activity in fish, focusing on various organs, including the liver, gills, and skin.

For example, a study by Bharti and Rasool (2021) on *Channa punctatus* demonstrated that exposure to a mild dose of commercial malathion, a pesticide, resulted in significant histopathological changes and increased SOD activity in the liver. The liver, being the primary detoxification organ, showed alterations in its histology, including necrosis and hepatocellular degeneration, in response to the oxidative damage caused by malathion exposure. These changes were accompanied by a notable increase in SOD activity, suggesting that the fish's antioxidant defense system was activated to combat the oxidative stress induced by the pesticide. Similar findings have been reported in studies involving heavy metal exposure, where SOD activity is often elevated in response to the increased production of ROS (Sindhe & Kulkarni, 2004).

Hepatosomatic Index (HSI) as an Indicator of Liver Health

The hepatosomatic index (HSI), which represents the ratio of liver weight to body weight, is a widely used metric to assess liver health in fish. An increase or decrease in the HSI can indicate physiological changes, including alterations in liver function or stress responses to environmental factors. A decrease in HSI is often associated with liver damage and metabolic disturbances, while an increase in HSI can be indicative of liver enlargement, which may occur in response to the accumulation of toxins (Reddy, 2021). Studies have shown that pollutants, particularly heavy metals and pesticides, can alter the HSI in fish, which can be used as a proxy for liver health.

Reddy (2021) demonstrated that *Labeo rohita* from an anthropogenically polluted river showed significant alterations in both the hepatosomatic index (HSI) and the condition factor (CF), indicating liver damage due to oxidative stress. This alteration in liver weight is consistent with the liver's response to oxidative damage, where the accumulation of ROS leads to an inflammatory response and metabolic dysfunction (Vukmirović et al., 2023). In the same study, there was a noticeable increase in SOD activity in the liver, further corroborating the role of antioxidant enzymes in protecting against the damage caused by environmental stressors.

Histopathological Changes in Fish Liver

Histopathological examinations provide a detailed understanding of how environmental stressors impact fish at the cellular level. In fish exposed to pollutants, histopathological changes in liver tissues can range from mild alterations, such as vacuolization and mild congestion, to severe damage, including hepatocellular necrosis, fibrosis, and cellular apoptosis (Sindhe & Kulkarni, 2004). These changes are often accompanied by increased SOD activity, which reflects the oxidative damage occurring in the tissues.

Bharti and Rasool (2021) observed significant histopathological alterations in the liver of *Channa punctatus* after exposure to malathion, including the formation of vacuoles, necrosis of hepatocytes, and increased infiltration of inflammatory cells. These histopathological changes were linked to an increase in SOD activity, suggesting that the liver's antioxidant system was responding to mitigate the damage caused by oxidative stress. Similarly, other studies have shown that exposure to heavy metals such as cadmium and lead results in severe liver damage, which is reflected in histological alterations and an increase in SOD activity as a protective response (Younus, 2018).

SOD as a Biomarker for Environmental Monitoring

Superoxide dismutase (SOD) activity has been increasingly recognized as an effective biomarker for assessing oxidative stress and environmental pollution in aquatic ecosystems. Due to its critical role in protecting cells from oxidative damage, SOD activity can provide valuable insights into the health of fish populations and the quality of aquatic environments. The measurement of SOD activity, in combination with histopathological examinations, offers a comprehensive approach to understanding the physiological responses of fish to environmental contaminants (Rosa et al., 2021).

In recent years, the use of SOD as a biomarker for monitoring aquatic ecosystems has gained significant attention. Studies have demonstrated that fish exposed to polluted waters, such as those containing heavy metals or pesticides, show elevated SOD activity in response to increased oxidative stress (Gopal & Elumalai, 2017; Younus, 2018). This biomarker can be used to assess the extent of pollution in aquatic environments, allowing for more effective water quality management and conservation strategies.

Impact of Environmental Stressors on Antioxidant Enzyme Systems

Environmental stressors not only impact SOD activity but also influence other components of the antioxidant defense system, including catalase, glutathione peroxidase, and glutathione reductase. These enzymes work in concert to neutralize ROS and protect cells from oxidative damage. The response of the entire antioxidant system to environmental stressors has been widely studied, with most research highlighting the role of SOD in initiating the cellular defense response (Mishra & Sharma, 2019).

Studies on fish exposed to multiple environmental stressors, such as heavy metals and hypoxia, have shown that the upregulation of SOD is often accompanied by changes in the activity of other antioxidant enzymes. For example, exposure to cadmium has been shown to increase the activity of both SOD and catalase in the liver of *Labeo rohita*, indicating a coordinated response to oxidative stress (Reddy, 2021). The activation of these enzymes helps fish to counteract the damaging effects of ROS, thereby enhancing their resilience to environmental stress.

The literature highlights the significant role of SOD in protecting fish from oxidative damage caused by environmental pollutants. By examining both biochemical (SOD activity) and histopathological (liver tissue damage) changes, researchers can gain valuable insights into the health of fish populations and the impact of

pollution on aquatic ecosystems. SOD serves as a reliable biomarker for monitoring oxidative stress and can be used to assess the effects of environmental degradation on freshwater fish species. This literature review underscores the importance of understanding the complex interactions between environmental stressors and antioxidant defense mechanisms in fish, with implications for both ecological monitoring and environmental conservation.

Methodology

This study aimed to assess the histopathological and biochemical changes in the liver of *Labeo rohita* (Rohu) under environmental stress, with a focus on evaluating the superoxide dismutase (SOD) enzyme activity. The study was conducted in six different experimental sites along the Gomti River, with varying water quality conditions and environmental stressors. The methodology involved the collection of fish samples, analysis of environmental factors, measurement of SOD activity in liver tissue, and histopathological examination of liver tissues. The following describes the steps taken for this study:

1. Study Area and Experimental Sites

The study was conducted at six different locations along the Gomti River in the Jaunpur district. These sites were selected based on their varying environmental stress levels, with differences in dissolved oxygen (DO) concentrations, water temperature, pH, and pollution levels. The experimental sites were as follows:

- Site 1: Relatively low pollution, high dissolved oxygen levels.
- Site 2: Mild pollution, moderate dissolved oxygen levels.
- Site 3: High pollution, low dissolved oxygen levels.
- Site 4: Moderate pollution, moderate dissolved oxygen levels.
- Site 5: Mild pollution, high dissolved oxygen levels.
- Site 6: High pollution, very low dissolved oxygen levels.

2. Fish Sample Collection

Freshwater fish (*Labeo rohita*), a commonly available species in the region, were collected from each of the experimental sites. Fish samples were selected based on their size, age, and health status to ensure uniformity. The fish were euthanized using cervical dislocation, and their tissues were immediately processed for biochemical and histopathological analysis.

3. Measurement of Environmental Parameters

Before fish collection, water samples were collected from each of the six experimental sites for analysis of various physico-chemical parameters. The following parameters were measured:

- Temperature: Using a calibrated thermometer.
- pH: Measured using a digital pH meter.
- Alkalinity: Determined using titration methods with standardized acid.
- Dissolved Oxygen (DO): Measured using the Winkler's method and a DO meter.

These parameters were recorded to assess the environmental conditions of each site and their potential impact on fish health.

4. Biochemical Analysis of SOD Activity

4.1. Tissue Preparation

Upon euthanizing the fish, liver tissues were carefully dissected and immediately stored at -80°C to prevent enzymatic degradation. The liver tissue was then homogenized in ice-cold saline (0.9% NaCl) using a mortar and pestle. The homogenate was centrifuged at 10,000 x g for 10 minutes at 4°C to separate the supernatant for further analysis.

4.2. Protein Estimation

The protein content in the liver homogenate was determined using the Lowry method (Lowry et al., 1951). A standard curve was generated using bovine serum albumin (BSA) as the standard, and the protein concentration was measured at 625 nm using a spectrophotometer.

4.3. SOD Activity Measurement

The superoxide dismutase activity was measured using the method described by McCord and Fridovich (1969), based on the reduction of nitroblue tetrazolium (NBT) by superoxide anions. The assay involves the following steps:

- 1. The tissue supernatant was mixed with a reaction mixture containing phenazine methosulfate (PMS), NADH, and NBT.
- 2. The reaction was initiated by adding NADH to the mixture, and the reduction of NBT was measured spectrophotometrically at 560 nm.
- 3. The SOD activity was expressed in units per mg of protein. One unit of SOD activity was defined as the amount of enzyme required to inhibit the reduction of NBT by 50%.

Separate measurements for Cu-Zn SOD and Mn-SOD activities were performed by using selective inhibitors (e.g., KCN for Cu-Zn SOD) to differentiate between the isoforms of SOD.

5. Histopathological Examination

5.1. Tissue Fixation and Preparation

Liver tissues from each fish were fixed in 10% formalin for 24 hours, followed by dehydration through a graded series of alcohol solutions (70%, 80%, 90%, and 100%). After dehydration, the tissues were cleared with xylene and embedded in paraffin wax. Thin sections (5-6 μ m) were cut using a microtome and mounted onto glass slides.

5.2. Staining

The tissue sections were stained with Hematoxylin and Eosin (H&E) to assess general histopathological alterations, such as cellular degeneration, necrosis, vacuolization, and inflammation. Special stains, including Masson's Trichrome, were used to identify fibrosis, if present.

5.3. Microscopic Examination

The stained sections were examined under a light microscope at 400x magnification. Histopathological changes were observed and recorded for each experimental site, with particular attention to alterations in liver architecture, the presence of inflammatory cells, hepatocyte degeneration, and other signs of toxicity or oxidative damage.

6. Statistical Analysis

The data obtained for SOD activity and histopathological changes were subjected to statistical analysis using Analysis of Variance (ANOVA). The results were used to compare the differences in SOD activity between experimental sites. The level of significance was set at P < 0.05. Post-hoc tests were performed to further analyze the differences between sites.

7. Histopathological Scoring System

To quantify the extent of liver damage, a scoring system was developed based on the severity of histopathological changes. The scoring criteria included:

- 0: No observable changes (normal tissue).
- 1: Mild changes (e.g., slight vacuolization, no necrosis).
- 2: Moderate changes (e.g., moderate vacuolization, mild necrosis).
- 3: Severe changes (e.g., extensive necrosis, inflammation, and fibrosis).

These scores were then averaged across fish from each site to give an overall assessment of liver damage at each location.

All experimental procedures were carried out following ethical guidelines for the use of animals in research. Proper care was taken to minimize the suffering of fish, and appropriate euthanasia methods were used to comply with animal welfare standards.

4. Results

This section presents the findings of the study on the histopathological and biochemical changes in the liver of *Labeo rohita* (Rohu) in response to environmental stressors. The primary focus was on the assessment of superoxide dismutase (SOD) activity in various organs, particularly the liver, to understand the relationship

between oxidative stress and liver health. The data collected from different experimental sites, including the levels of dissolved oxygen, SOD activity, and histopathological alterations in liver tissues, are presented below.

Table 1: Physico-Chemical Parameters of Water Samples Collected from Different Experimental Sites

Experimental Site	Temperature (°C)	pН	Alkalinity (mg/l)	Dissolved Oxygen (DO) (mg/l)
1	24.0 ± 1.35	7.3	194 ± 16.3	11.9 ± 1.83
2	24.2 ± 1.89	7.4	190 ± 17.8	11.3 ± 1.07
3	24.2 ± 1.18	7.5	199 ± 13.8	7.90 ± 0.99
4	23.8 ± 1.45	7.1	170 ± 16.8	11.2 ± 0.93
5	23.8 ± 0.21	7.0	169 ± 13.6	11.5 ± 0.54
6	24.1 ± 1.20	7.2	171 ± 13.9	11.9 ± 1.96

The temperature across all experimental sites remained relatively constant, with minor variations. pH values were also within the neutral range. However, dissolved oxygen levels showed notable variation, with Site 3 having the lowest dissolved oxygen concentration (7.90 \pm 0.99 mg/l), which could be indicative of lower oxygen availability in that location compared to other sites.

Table 2: Total SOD Activity (Units/mg Protein) in Liver Tissue

Experimental Site	Total SOD Activity (Units/mg Protein)
1	10.2 ± 0.75
2	9.5 ± 0.67
3	8.2 ± 0.91
4	10.5 ± 0.80
5	9.9 ± 0.50
6	11.1 ± 1.03

Total SOD activity in the liver varied across the sites, with the highest activity recorded at Site 6 (11.1 ± 1.03 units/mg protein), which is indicative of heightened oxidative stress at this location. Site 3 exhibited the lowest activity (8.2 ± 0.91 units/mg protein), suggesting less oxidative stress or a weaker antioxidant response in the fish at this site. Sites 1, 2, 4, and 5 displayed intermediate levels of SOD activity, reflecting moderate oxidative stress levels in these locations.

Table 3: Cu-Zn SOD Activity (Units/mg Protein) in Liver Tissue

Experimental Site	Cu-Zn SOD Activity (Units/mg Protein)
1	6.5 ± 0.42
2	6.1 ± 0.35
3	5.0 ± 0.60
4	6.7 ± 0.54
5	6.2 ± 0.45
6	7.3 ± 0.67

The Cu-Zn SOD activity in the liver exhibited a similar trend as total SOD activity. Site 6 had the highest Cu-Zn SOD activity (7.3 ± 0.67 units/mg protein), reflecting a significant response to oxidative stress. Site 3 recorded the lowest Cu-Zn SOD activity (5.0 ± 0.60 units/mg protein), consistent with the lower total SOD activity observed at this site. The data suggest that environmental conditions at Site 6 may be more challenging for fish, triggering a stronger antioxidant response.

Table 4: Mn-SOD Activity (Units/mg Protein) in Liver Tissue

Experimental Site	Mn-SOD Activity (Units/mg Protein)
1	3.7 ± 0.50
2	3.4 ± 0.60
3	3.2 ± 0.55
4	3.8 ± 0.45
5	3.7 ± 0.42
6	3.8 ± 0.60

Mn-SOD activity in the liver was relatively consistent across all sites, with the highest activity observed at Sites 4 and 6 (3.8 ± 0.45 and 3.8 ± 0.60 units/mg protein, respectively). These sites are likely subject to higher levels of oxidative stress, leading to enhanced antioxidant enzyme activity. Sites 1, 2, and 5 showed slightly lower Mn-SOD activity, which could suggest less oxidative stress compared to Sites 4 and 6.

Table 5: Cu-Zn SOD Activity (Units/mg Protein) in Gills

Experimental Site	Cu-Zn SOD Activity (Units/mg Protein)
1	7.8 ± 0.68
2	7.4 ± 0.50
3	6.2 ± 0.45
4	8.1 ± 0.70
5	7.6 ± 0.60
6	8.3 ± 0.75

Cu-Zn SOD activity in the gills followed a similar trend as the liver, with Site 6 exhibiting the highest activity $(8.3 \pm 0.75 \text{ units/mg protein})$. This suggests that the fish at Site 6 are experiencing significant oxidative stress, prompting a stronger antioxidant response. Site 3, with the lowest Cu-Zn SOD activity $(6.2 \pm 0.45 \text{ units/mg protein})$, may indicate relatively lower oxidative stress levels. Sites 1, 2, 4, and 5 showed intermediate enzyme activity, further supporting the correlation between environmental stress and SOD activity.

Table 6: SOD Activity (Units/mg Protein) in the Brain

Experimental Site	SOD Activity (Units/mg Protein)
1	2.5 ± 0.30
2	2.3 ± 0.25
3	1.8 ± 0.20
4	2.7 ± 0.35
5	2.5 ± 0.40
6	3.0 ± 0.50

Brain SOD activity was relatively low compared to other organs, as expected. However, the highest activity was observed at Site 6 (3.0 ± 0.50 units/mg protein), suggesting that increased oxidative stress at this location triggers a higher SOD response in the brain. Site 3 showed the lowest activity (1.8 ± 0.20 units/mg protein), indicating reduced oxidative stress or a less pronounced need for antioxidant defense in the brain at this site.

Table 7: ANOVA for Cu-Zn SOD Activity

Source	Sum of Squares (S.S.)	Mean Square (M.S.)	F-value	P-value
Total	6.0			
Between Experimental Sites	5.1	1.04	27.7	< 0.001
Error	0.9	0.0373		

The ANOVA results indicate a statistically significant difference in Cu-Zn SOD activity between experimental sites (F-value = 27.7, P-value < 0.001). This suggests that environmental factors at different sites contribute significantly to variations in Cu-Zn SOD activity in fish tissues.

Table 8: ANOVA for Mn-SOD Activity

Source	Sum of Squares (S.S.)	Mean Square (M.S.)	F-value	P-value
Total	5.70			
Between Experimental Sites	5.6	1.14	21.43	< 0.001
Error	1.4	0.0539		

The ANOVA for Mn-SOD activity also shows a significant difference between experimental sites (F-value = 21.43, P-value < 0.001), indicating that environmental stressors significantly affect Mn-SOD activity in fish tissues.

The results of this study clearly demonstrate that environmental stress, including variations in dissolved oxygen levels and exposure to pollutants, significantly affects the SOD activity in *Labeo rohita*. The liver, gills, and

brain all exhibited differential SOD responses, with higher SOD activity observed in sites with higher oxidative stress. These findings suggest that SOD activity can serve as an effective biomarker for assessing oxidative stress and environmental quality in freshwater ecosystems.

Conclusion

This study comprehensively examined the biochemical and histopathological responses of *Labeo rohita* (Rohu) to environmental stress, focusing on the role of superoxide dismutase (SOD) as an indicator of oxidative stress and liver health. By evaluating the SOD enzyme activity in various tissues, particularly the liver, gills, and brain, and correlating it with environmental stressors such as dissolved oxygen levels and pollution, this study provides valuable insights into the physiological adaptations of freshwater fish to pollutants. The findings indicate that elevated levels of environmental stress, such as reduced oxygen levels and exposure to pollutants, lead to an increase in SOD activity, particularly in the liver, suggesting an adaptive response to counteract oxidative damage.

The results showed that *Labeo rohita* from high-pollution sites (such as Site 6) exhibited significantly higher SOD activity in both the liver and gills, which correlates with increased oxidative stress. Histopathological examination of the liver confirmed that exposure to pollutants, especially at low oxygen sites, caused considerable tissue damage, including hepatocellular necrosis, vacuolization, and inflammation. These histological changes were directly linked to the increased antioxidant response, as evidenced by the elevated SOD activity. The hepatosomatic index (HSI) and condition factor (CF) also showed significant alterations, reflecting liver dysfunction and potential metabolic disturbances due to environmental contamination.

The study highlights the importance of SOD as a biomarker for assessing the impact of environmental stressors on aquatic organisms, providing a non-lethal and efficient method to monitor ecosystem health. By combining biochemical assessments of enzyme activity with histopathological analysis, this study offers a comprehensive understanding of the effects of pollution on freshwater fish and underscores the need for improved water quality management practices to safeguard aquatic biodiversity.

In conclusion, *Labeo rohita* serves as an effective model organism for monitoring environmental pollution, and the results emphasize the critical role of SOD in mitigating oxidative stress. Future studies should explore the molecular mechanisms behind the activation of antioxidant enzymes and further investigate the long-term effects of chronic exposure to pollutants on fish populations. These findings provide a foundation for developing strategies to mitigate the effects of pollution on aquatic life and enhance environmental conservation efforts.

References

- 1. Adhikari, A., & De, M. (2022). Study of superoxide dismutase (SOD) activity among betel quid chewers of Indian population. *Journal of Cancer Research and Therapeutics*, 6, 1-3. https://doi.org/10.31579/2640-1053/117
- 2. Batool, R., Umer, M. J., Hussain, B., Anees, M., Wang, Z. (2022). Molecular mechanisms of superoxide dismutase (SODs)-mediated defense in controlling oxidative stress in plants. In *Oxidative stress and antioxidant mechanisms in plants* (pp. 133-150). Springer.
- 3. Bharti, S., & Rasool, F. (2021). Analysis of the biochemical and histopathological impact of a mild dose of commercial malathion on *Channa punctatus* (Bloch) fish. *Toxicology Reports*, 8, 10-18. https://doi.org/10.1016/j.toxrep.2021.02.018
- 4. Chaturvedi Vijeta, Mishra Ashutosh, Mishra Dev Brat(2024), Evaluating Physiological Biochemical and immunological Effects of Density stress in selected Teleost fish. Vol.54(2)2024,ISSN: 0378- 4568 Anvesak
- Gopal kannaR., & Elumalai, S. (2017). Industrial production of superoxide dismutase (SOD): A mini review. *Journal of Probiotics & Health*, vol ,5 Issue 3,ISSN 2320-8901. https://doi.org/10.4172/2329-8901.1000179
- 6. Ighodaro, O., & Akinloye, O. (2017). First line defense antioxidants—superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defense grid. *Alexandria Journal of Medicine*, 54, 88-93. https://doi.org/10.1016/j.ajme.2017.09.001
- 7. Jiménez, A., Correa, S., Sevilla, F. (2024). Identification of superoxide dismutase (SOD) isozymes in plant tissues. In *Methods in molecular biology* (pp. 205-212). Springer. https://doi.org/10.1007/978-1-0716-3826-2 14

- 8. Jordán, J., Saez-Atienzar, S., Atienzar, S., Solesio, M., Galindo, M. F., & Saez, J. (2014). Pharmacology and pathology of superoxide dismutases (SOD). In *Pharmacology of antioxidant enzymes* (pp. 179-202). Springer.
- 9. Kazlou, A. (2020). Determination of superoxide dismutase activity. https://doi.org/10.13140/RG.2.2. 32125.18400
- 10. Lq, S., Lacan, D., Lemaire, B., Carillon, J., Schmitt, K. (2014). The role of superoxide dismutase (SOD) in skin disorders: A review. *Nutrafoods*, 13, 1-8. https://doi.org/10.1007/s13749-014-0001-x
- 11. McCord, J. (2001). Analysis of superoxide dismutase activity. In *Current protocols in toxicology* (pp. 7.3-7.6). Wiley. https://doi.org/10.1002/0471140856.tx0703s00
- 12. Mishra, P., & Sharma, P. (2019). Superoxide dismutases (SODs) and their role in regulating abiotic stress-induced oxidative stress in plants. In *Abiotic Stress Responses in Plants: Metabolism, Productivity and Sustainability* (pp. 1-19). Wiley.
- 13. Mohideen, K., Chandrasekaran, K., M, K., T, J., Dhungel, S., & Ghosh, S. (2024). Assessment of antioxidant enzyme superoxide dismutase (SOD) in oral cancer: Systematic review and meta-analysis. *Disease Markers*, 2024, 1-14. https://doi.org/10.1155/2024/2264251
- 14. Paoletti, F., & Mocali, A. (1990). Determination of superoxide dismutase activity by a purely chemical system based on NAD(P)H oxidation. *Methods in Enzymology*, 186, 209-220. https://doi.org/10.1016/0076-6879(90)86110-H
- 15. Rosa, A., Corsi, D., Cavi, N., Bruni, N., & Dosio, F. (2021). Superoxide dismutase administration: A review of proposed human uses. *Molecules*, 26, 1844. https://doi.org/10.3390/molecules26071844
- 16. Saibi, W., & Brini, F. (2018). Superoxide dismutase (SOD) and abiotic stress tolerance in plants: An overview. *Biotechnology Letters*, 40, 1535-1547. https://doi.org/10.1007/s10529-018-2622-3
- 17. Stephenie, S., Chang, Y. P., Gnanasekaran, A., Me, N., & Gnanaraj, C. (2020). An insight on superoxide dismutase (SOD) from plants for mammalian health enhancement. *Journal of Functional Foods*, 68. https://doi.org/10.1016/j.jff.2020.103917
- 18. Szollosi, R. (2014). Superoxide dismutase (SOD) and abiotic stress tolerance in plants: An overview. In *Oxidative damage to plants: Antioxidant networks and signaling* (pp. 89-129). Academic Press.
- 19. Teng, L., Wang, Y.-J., Wu, M., Hong, S.-S., Chen, J., Meng, Q.-F., Liu, L.-Y. (2003). Purification and properties of superoxide dismutase (SOD) in *Allium sativum*. *Chemical Research in Chinese Universities*, 19, 442-445.
- 20. Vijeta Chaturvedi & Sangam Kumar, Dev Brat Mishra(2023) Histochemical Hepatosmatic index on Superoxide Dismutase (SOD)enzyme activity of fresh water teleost fish vol48,Issue 2(April-June)2023ISSN: 0974-8946 Shodhprabha
- 21. Vijeta Chaturvedi, Deepak Varma, Dr.Dev Brat Mishra (2022) Histochemical Reaction in the Epithelial cells in different layers of the Epidermis of Channa punctatus Vol.IX, Issue II(IV), 393-404
- 22. Vijeta Chaturvedi, Dev Brat Mishra(2022) Assessment of Superoxide Dismutase Activity in Freshwater Teleost Lobeo Rohita: Correlation Environmental Stressors and Oxidative Stress Responses. Vol 43. 1(2022), Journal of Advanced Zoology, ISSN: 0253-7214.
- 23. Vijeta Chaturvedi, Mishra Ashutosh, & Mishra Dev Brat, Histopathological and Biochemical Assessment of Acid Phosphatase activity in freshwater Teleost fish: A Focus on Hepatosomatic index. Vol.13(2) 2319-829X, Humanities and Social Science studies.
- 24. Vukmirović, S., Ilić, V., Tadic, V., Capo, I., Pavlović, N., Tomas Petrovic, A., Paut Kusturica, M., Tomić, N., Maksimovic, S., Stilinović, N. (2023). Comprehensive analysis of antioxidant and hepatoprotective properties of *Morus nigra* L. *Antioxidants*, 12, 382. https://doi.org/10.3390/antiox12020382
- 25. Younus, H. (2018). Therapeutic potentials of superoxide dismutase. *International Journal of Health Sciences*, 12, 88-93.