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## Effect Of Refrigerating Temperature On Protein Compositional Level In Body Muscle Of Labeo Bata (Hamilton)

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#### Abstract

A 14 days experiment was conducted to analyze the changes of protein level in *Labeo bata* (Hamilton 1822) due to refrigeration. The result indicates that the proportion of protein in the muscle of fish decreased with the increased number of days the fish in the refrigerator. Due to refrigerating temperature the muscle got tighter with the number of days increased and the protein content decreased. Protein composition was classified into five fractions based on solubility, including water-soluble proteins, NPN, salt-soluble proteins, alkali-soluble proteins and alkali-insoluble proteins. Effects of heating cooling on the content of each fraction With the decrease of temperature, significant changes in content of water-soluble proteins, salt-soluble proteins and alkali-soluble proteins were observed, and little changes for NPN and alkali-insoluble proteins.

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Keyword: Labeo bata, protein, refrigeration, fish muscle

## 1. Introduction

Fish is a good source of high quality protein, vitamin and essential minerals. India is one of the largest producers of freshwater fish in the world. *Labeo bata* is one of the major species produced in India and Bangladesh and has a unique taste and texture, and is very popular among consumers especially in Assam, west Bengal and Odisa, as well as other part of the country. Protein is the main nutritional ingredient of fish muscle (meat) and the main supporting composition of muscle structure, and the changes of protein will directly affect the structure, textural and sensory properties of fish meat.

Infrigidation is one of the major steps in preservation of fish. Difference chilling temperature can cause varying degrees of degeneration or degradation of protein in the muscle, leading to the differences in organizational characteristics, texture and sensory quality (Palka and Daun, 1999). It is reported that the stability of protein during chilling is close related to its structure, changes in texture, flavor, color and other sensory qualities, also the protein solubility and collagen content of various muscles, including beef, pork, chicken, rabbit, shrimp, sea fish, etc. few studies involved in the changes of protein composition in fresh water fish chilled at different temperatures. The results obtained from different specie muscles could have something to do with the muscle type and structure, and cooling properties of protein, such as the protein compositions and solubility, as well as the content and nature of the intramuscular collagen.

The effects of cooling temperature on the texture, color of *Labeo bata* muscle were studied, and the objective of this paper was to discuss the effects of chill treatments on the protein components of fish muscle, and further understand the relationship between the texture and protein composition. In addition, the biochemical characteristics changes of collagen in *Labeo bata* carp during chilling were studied by Bradford Reagent.

Labeo bata is predominantly a bottom feeder and herbivorous fish (Chondar, 1999). Datta et al. (1986) reported that Labeo bata can grow up to 44-67g in 10-12 months in paddy field with other carps. Datta et al. (1996) also Available online at: <a href="https://iazindia.com">https://iazindia.com</a> 340

observed the growth of *Labeo bata* from 0.09-0.2 g to 23-38 g in 6-10 months under mono culture with sewage water as input.

Minor carp *Labeo bata* (pictures: 1) is a freshwater subtropical species which is commonly known as 'Bangon Bata'. This fish is commercially important and target species for commercial small- and large-scale fishers in Bangladesh, India and Pakistan. It is also used by both culture and capture fisheries nowadays. L. bata is in great demand in the market because of its high nutritional value and good taste (Bhuiyan, 1964). This fish contains about 15.42% of protein and 3.73% of lipid (Ahmed et al., 2012).

#### 2. Materials and methods

Sample treatments Labeo bata weighing between (35-47gm) was bought from Nagaon Bara Bazar (since the fish was live and brought in a small glass fish pot). Then it was transferred to the laboratory, the fish were beheaded, gutted and washed. Before it was killed muscle of that live fish was isolated and then the fish killed and the skin was removed and the muscles were trimmed of visible fat and connective tissue. The muscles were cut with the fibers parallel to the longest axis. The fish blocks were individually weighed and packed within the centrifuged tube and stored in the laboratory fridge.

#### 2.1 Chemical preparation

The Bradford reagent consists of the dye Coomassie Brilliant Blue G in phosphoric acid and methanol or ethanol.

Brilliant Blue G forms a complex with proteins in solution resulting in a shift in the absorption maximum of the dye from 465 to 595nm. The absorption is proportional to the amount of protein present. This reagent can be used to quantify proteins in the concentration range from 0.1 to approximately 1.4 mg/ml. The Bradford assay is compatible with reducing agents such as dithiothreitol; however it is only compatible with very low concentrations of detergents. If the samples contain detergent then the BCA assay is recommended. It is always advisable to prepare the standard in the same buffer as the sample to minimize any interference effects. The dye reagent reacts primarily with arginine residues and less so with histidine, lysine, tyrosine, tryptophan, and phenylalanine residues. Compared to "average" proteins, the Bradford assay is more sensitive to bovine serum albumin, by a factor of about two Immunoglobulin G (IgG - gamma globulin) is therefore the preferred protein Standard.

## 2.2 Materials required

**Bradford Reagent** 

Suitable tubes to hold and mix 3.1ml samples

Plastic disposable cuvettes (Jenway 060 084)

Standard protein solution of known concentration (2Mg/ml)

Method

Preparation of the Bradford reagent

The Bradford reagent is available commercially from a number of different sources. However it can be Prepared as follows:

- 1. Dissolve 100mg Coomassie Brilliant Blue G-250 in50ml 95% ethanol; add 100ml 85% (w/v) Phosphoric acid.
- 2. Once the dye has completely dissolved, dilute to 1 litre with deionised water.
- 3. Filter through Whatman #1 paper just before use.

## 2.3 Flow Chart Showing The Process The Process Of Extraction

From fish 1gm tissue is weight and weight and prepared (1gm tissue in 2ml of distilled water

From the above solution 1ml solution is taken

1ml cold 10% TCA is added

Solution is then centrifuged at about 4000rpm for 5 -10 minute

Supertant is then decanted

2.5 ml of 5% TCA is added to it and centrifuged

Precipitated is then washed with 2.5 ml of alcohol and centrifuged

Again wash with alcohol ether (3:1 and centrifuged

Then after tissue is inverted in a stand having a tissue paper below.

After 30 minutes 3 ml of 0.1 N NaoH is added and left in water bath at 37° FOR 3-4 hours

Supertant is warmed up

Available online at: <a href="https://jazindia.com">https://jazindia.com</a>

 $20\mu l(10\mu g),40\mu l(20\mu g),60\mu l(30\mu g),80\mu l(40\mu g)$  of 0.1 mg/ml

BSA were taken out separate cuvettes and made the volume  $100\mu l$  with distilled water  $100\mu l$  tissue extracted is taken as unknown. The cuvette with no protein standard in it served as blank

1.5 ml of Bradford reagent is added to each cuvette

Cuvette are covered with plastic paraffin film

0.1% Stock solution and mixed well

Then the cuvette were incubated at room temperature for 10 minutes

Absorbance of cuvette is measured at 595nm

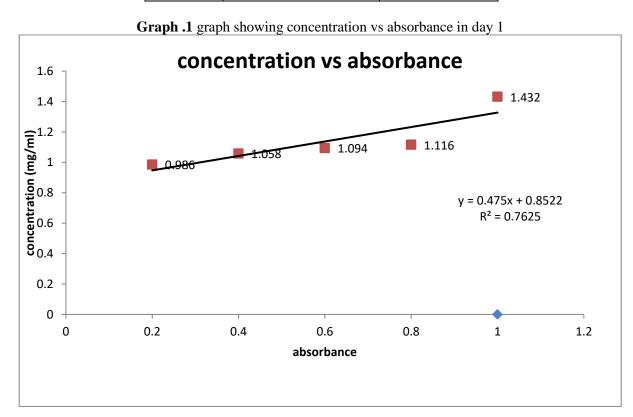
Absorbance of each BSA standard as function of its theoretical concentrated plotted

Centrifugation of unknown is calculated from the graph

#### 3. Result

**Table.** 1 showing concentration and absorbance parameters in day 1

sample	Concentration(mg/ml)	Absorbance <sub>595nm</sub>
Blank	1ml	0.00
S1	0.2(mg/ml)	0.986
S2	0.4(mg/ml)	1.058
S3	0.6(mg/ml)	1.094
S4	0.8(mg/ml)	1.116
S5	1.0(mg/ml)	1.432
Unknown	Unknown(mg/ml)	1.673



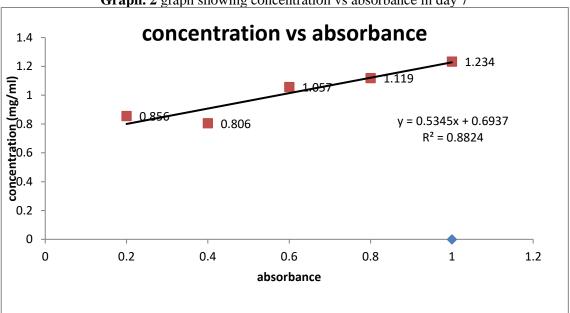
## Calculation

Unknown absorbance=1.637 Unknown concentration (mg/ml) =1.42 From the equation y = 0.475x + 0852

**Table. 2** showing concentration and absorbance parameters in day 7

sample	Concentration(mg/ml)	Absorbance <sub>595nm</sub>
Blank	1ml	0.00
S1	0.2(mg/ml)	.856
S2	0.4(mg/ml)	.806
S3	0.6(mg/ml)	1.057
S4	0.8(mg/ml)	1.119
S5	1.0(mg/ml)	1.234
Unknown	Unknown(mg/ml)	1.437

**Graph. 2** graph showing concentration vs absorbance in day 7



## Calculation

Unknown absorbance=1.437

Unknown concentration (mg/ml) =1.29

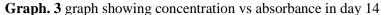
From the equation y = 0.534x + 0.693

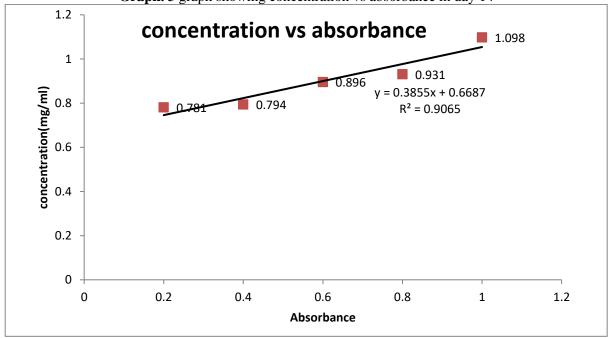
Therefore x = 1.117 mg/ml

Table. 3 showing concentration and absorbance parameters in day 14

sample	Concentration(mg/ml)	Absorbance <sub>595nm</sub>
Blank	1ml	0.00
S1	0.2(mg/ml)	.781

S2	0.4(mg/ml)	.794
S3	0.6(mg/ml)	.896
S4	0.8(mg/ml)	.931
S5	1.0(mg/ml)	1.098
Unknown	Unknown(mg/ml)	1.112





#### Calculation

Unknown absorbance=1.112 Unknown concentration (mg/ml) =1.098 From the equation y = 0.385x + 0.668Therefore x = 1.116 mg/ml

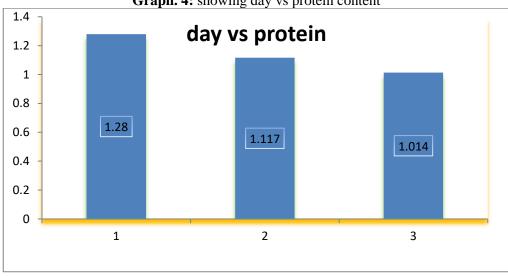
## 4. DISCUSSION

As shown in the above table -1, 2, and 3 variation of total protein content (mg/ml) in body the muscles of selected fishes (*Labeo bata*) collected from Nagaon Bara Bazar, dated 12.03.2020. The total protein content in body muscle of selected fish ranges from 1.280±1.114(mg/ml).

The highest protein content in the selected fish was 1.280±.314 and the lowest was 1.114±.093.

TABLE 4: showing protein content in three different days

Sample	day	proteir
1	1	1.28
2	7	1.117
3	14	1.014



**Graph. 4:** showing day vs protein content

#### 5. CONCLUSION

Total protein content in body muscles of Labeo bata was higher on the 1st day of experiment and lower in the last of experiment i.e. on 14th day. At the very first day the fish muscle content highest amount of protein i.e. ±1.28 (mg/ml) and then the fish was kept under observation for 7 days and the same process repeated and protein content of the fish was calculated. On the 7<sup>th</sup> day the fish muscle contain slightly less protein then the  $1^{\text{st}}$  day i.e.  $\pm 1.117$  (mg/ml). On the  $14^{\text{th}}$  day same process was repeated and protein content was measured at  $\pm 1.116$  (mg/ml).

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