

Journal of Advanced Zoology

ISSN: 0253-7214 Volume 43 Issue S 1 Year 2022 Page 544-448

A Preliminary Screening And Determination Of Biochemical Properties Of Bacterial Isolates Producing An Anticancerous Enzyme Asparaginase From Some Sewage Water Samples Of Bhopal, M.P. India

Shreya Tiwari^{1*}, Rakesh Mehta², Ragini Gothalwal³, Laxmikant Pandey⁴

1*Research scholar Department of Biotechnology Barkatullah University Bhopal
2Principal and Head Department of botany and Biotechnology, GOVT MGM College Itarsi
3Head department of biotechnology Barkatullah university Bhopal
4Head department of Biotechnology St Aloysius College Jabalpur

*Corresponding Author: Shreya Tiwari

*Research scholar Department of Biotechnology Barkatullah University Bhopal, - Shreyat714@gmail.com

Abstract:

Microorganisms have had a major impact on the development of medical science since the discovery that they not only cause infections but also produce certain organic compounds that cure infections and help treat a variety of non-infectious diseases Though, microbes are ubiquitous, but their metabolic capabilities are greatly influenced by the habitat they survive with unique conditions of pH, temperature, pressure, oxygen, light, nutrients and salinity, there is a high efficiency for those to yield metabolites to exhibit special biological activities The production of enzymes is a pursuit central to the modern biotechnology industry. Asparaginase is one such important enzyme finds their use in the pharmaceutical, biosensor and food industries and has anticarcinogenic potential for the treatment of acute lymphoblastic leukemia, lymphomas and other cancers. Asparaginase are naturally occurring enzymes expressed and produced by animal tissues, bacteria, plants, and in the serum of certain rodents, but not in mankind. Erwinia carotovora, Pseudomonas stutzeri, Pseudomonas aeruginosa, E. coli etc has been known to produce L-asparaginase, though its commercial production is restricted to use of Erwinia chrysanthemi and E. coli as per the latest information. Thus the present work was intended to screen out certain new bacterial isolates with in vitro asperginase producing potential from sewage water sources. The sewage water samples were collected from 5 different location of Bhopal and were subjected to brain heart infusion media containing amino acid asparagine and phenol red indicator. The bacterial isolates forming pink or red colonies or hallow around indicating asparaginase producing capability were picked and pure cultured. The three asparaginase positive isolates one from M.P. Nagar & other two from AIIMS sewage sample reported to be bacillus and coccus respectively on Gram's staining which when subjected to biochemical characterization using catalase test, IMVC test and carbohydrate fermentation tests does not matches with the biochemical characteristics of standard E.coli and S. aureus. The outcomes of this preliminary screening encourages to further investigate the sewage water samples to screen out the possibly new & efficient asparaginase producing isolates with complete biochemical & molecular characterization in view of commercial prospects.

CC License CC-BY-NC-SA 4.0

Keywords: L-asparaginase, anticarcinogenic, enzyme production, wastewater

INTRODUCTION

Asparaginase is one such important enzyme finds its used in the pharmaceutical, biosensor and food industries and has anticarcinogenic potential against acute lymphoblastic leukemia, lymphomas and other cancers ¹⁻³. Asparaginase enzyme explicitly focuses the metabolism of cancer cells by manipulating deficits in metabolic pathways and catalyzing the collapse of L-asparagine into ammonia and L-aspartic acid, triggering nutrient starvation of cancer cells and bringing about their demise ⁴.

Microorganisms such as bacteria and fungi are promising sources for structurally diverse and potent bioactive compounds ^{5,6}. Enzymes are chiefly sourced from microorganism since in a short span to time they could be culture in huge masses where prospects of genetic manipulation of bacterial cells could enhances their enzyme production potential. In addition, due to the active and stable nature, microbial enzymes have been in prime choice over enzymes of plant and animal origin ^{7,8}.

Pseudomonas aeruginosa, *Pseudomonas stutzeri*, *Escherichia coli*, *Erwinia carotovora*, and several other bacteria have been known to produce L-asparaginase ⁹⁻¹². But *E. chrysanthemi* and *E. coli* are being used for commercial production of two types of asparaginase enzymes for clinical used as per the latest information ¹³.

Under harsh and toxic environments, the enzyme producing capacity and growth of microorganisms itself gets retarded mostly. However, many microorganisms undergo likely physiological or genetic adaptations their by enabling them to survive and release enzyme under harsh conditions ^{7, 14}. Sometimes, the sewage water becomes a tough environment for microbes because of toxicity generated due pharmaceutical pollutants, disinfectants, chemical, dyes etc., and leads to survival of microbial population different metabolic potentials. The present preliminary investigation is intended to screen out such microbial isolates from local sewage sources with biopharmaceutically significant metabolic potential with reference to asparaginase enzyme.

MATERIALS & METHODS

Sample Collection:

For the screening and isolation of suspected bacteria, sewage water samples were used to collect in sterile containers and taken to the laboratory. The sewage water samples were collected from various locations of Bhopal namely M.P Nagar zone-II, Anna Nagar, Meera Nagar, Saket Nagar and Barkheda Pathani.

Media Preparation:

For screening of asparaginase producing bacteria a modified brain heart infusion broth (Himedia M210) media containing 10 gm/l amino acid asparagine, 6 gm/l K2HPO4, 4 ml/l MgSO4 2 ml/l CaCl2 gm/l 2.5 ml of 2.5% phenol red indicator and 15 gm/l agar was prepared ¹⁶. All the contents were mixed followed by adjusting the pH at 7. After autoclaving, the media was poured aseptically into sterile petri plates.

Screening of Asparaginase Producing Bacteria:

The water samples collected from different sources were subjected to serial dilution in sterile distilled water in aseptic conditions. 10^{-5} , 10^{-7} and 10^{-8} dilutions from series were used to raise the microbial cultures by spreading 0.1 ml of samples in separate culture plates. After incubation at 37°C for 24 hours in bacteriological incubator the bacterial colonies forming pink or red or hallow around indicating asparaginase producing capability were picked and pure cultured.

Biochemical Characterization:

The suspected asparaginase producing bacterial isolates were subjected to partial characterization through Gram's staining, and some biochemical tests like catalase test, indole test, methyl red reduction test, voges proskauer test, and citrate utilization test. The carbohydrate fermentation test with Sucrose, Fructose, Lactose, Dextrose and Mannitol was also performed.

In vitro Asparaginase Production & Assay:

50 ml of nutrient broth (Himedia MM244) containing 1% asparagine maintained at pH 7 was prepared in 125 ml conical flasks. After autoclaving, the media was inoculated with 1 ml of the culture of suspected

bacterial isolates and incubated at 37°C for 48 hours with intermittent shaking. After incubation, 2 ml of fermentum was centrifuged at 10000 rpm then supernatant was retained in new sterile microfuge tube. The activity of asparaginase was assayed by well diffusion method again on modified BHI medium containing 1% asparagin and 0.01% phenol red indicator. Wells were prepared on mediums of 6 mm diameter and 10 µl of fermentation extract from each isolates separately were poured aseptically in wells. After 24 hours of incubation at ambient temperature pink coloured zones were observed around the wells for any asparaginase activity due to release of ammonia ¹⁵⁻¹⁶.

RESULTS & DISCUSSION

Screening of Asparaginase-Producing Bacteria:

In terms of preliminary investigation for search of asparaginase enzyme producing bacteria sewage water samples were collected from 5 different locations of Bhopal which are M.P Nagar zone-II, Anna Nagar, Meera Nagar, Saket Nagar and Barkheda Pathani. The sample from M.P Nagar zone-II and Saket Nagar were reported to have bacterial colonies with red or pink colour or hallow around them due to the breakdown of asparagine amino acid present in the culture media. Sewage water both from M.P Nagar zone-II and Saket Nagar nears AIIMS were observed to contain heavy microbial load even at 10^{-7} and 10^{-8} dilutions indicates heavy load of untreated organic pollutants in sump. Though the isolates with positive asparaginase from these samples were taken forward for further confirmatory studies. Recently, Tomar *et al.*, (2019) ¹⁷ isolated asperaginase producing bacteria soil samples using M9 media plate; where, Bhat *et al.*, (2015) ¹⁸ reported *Salinicoccus* sp. as asparaginase production microbe from soil. Though, Tarulli *et al.*, (2013) ¹⁹ reported 21 bacterial isolates with L-asparaginase activity on plate method assay by pink colour zones.

Biochemical Characterization:

There were three different types of bacterial isolates were reported with asparaginase producing ability & were coded as Sp-1, Sp-2 and Sp-3. The three asparaginase positive isolates one from M.P. Nagar zone-II & other two from AIIMS sewage Saket Nagar sample reported to be bacillus, short bacillus & coccobacillus type respectively on Gram's staining which when subjected to biochemical characterization using catalase test, IMVC test and carbohydrate fermentation tests does not matches with the biochemical characteristics of E. coli (MTCC-1687) and S. aureus (MTCC-737). The results of the biochemical tests are mentioned in table 1.

Wastewater can carry many opportunistic pathogens like Enterobacter cloacae, Enterococcus faecalis, Escherichia coli, Klebsiella pneumoniae, Proteus vulgaris or Pseudomonas aeruginosa ²⁰. Clostridium perfringens, Legionella pneumophila and Mycobacterium tuberculosis like species were found to be common ²¹. Using a combination of FISH probing and 16S rDNA clone inserts, Plumb, et al., (2001) ²² revealed the methanogenic population of Methanobacterium and Methanospirillum species from wastewater containing food dyes degraded under sulfidogenic and methanogenic conditions. Liu et al.,

 $(2015)^{23}$, identified 18 species of genera Longilinea, Georgenia, Desulforhabdus, Thauera, Desulfuromonas and Arcobacter in the sewerage system which are responsible for methane fermentation of sewage sludge, facilitating decomposition of macromolecular organic matter into simpler compounds (Cyprowski, *et al.*, 2018) ²⁰. The present investigation has the provision for precise molecular identification of the isolates Sp-1, Sp-2 and Sp-3 in later studies.

Table 1: The observation of microscopy & biochemical tests for 3 asparaginase producing bacteria

S.N.Tests for Characterization		Bacterial Isolates			
		S-1	S-2	S-3	
1.	Gram's Staining	Gram	Gram	-ve Gram -ve	
		+ve	Short	Coccobacillus	
		Rods	Rods		
2.	Catalase test	+ve	+ve	+ve	
3.	Indole test	-ve	-ve	-ve	
4.	Methyl red reduction test	+ve	+ve	+ve	

5.	Voges Proskauer	· test	+ve	+ve	+ve	
6.	Citrate utilization test		-ve	-ve	-ve	
			Mannitol+ve	-ve	-ve	
			Lactose -ve	-ve	-ve	
7.	Carbohydrate	Fermentation	Sucrose +ve	-ve	-ve	
	Tests		Fructose -ve	-ve	-ve	
			Dextrose +ve	-ve	-ve	

In vitro Asparaginase Production & Assay:

The three bacterial isolates earlier screened for asparaginase activity when subjected to fermentation in broth containing amino acid asparagine, yields the asparaginase enzyme in medium during fermentation. The cell free fermentation extract when poured into well bored into same media with asparagine and phenol red indicator gives a clear pink zone. This indicates that all the three bacterial isolated have the ability to release asparaginase enzyme when allowed to grow in fermentation medium which encourages their possibilities explore microbes and their sources in wider perspective.

CONCLUSIONS

The treated or untreated wastewater gathers organic matter, variety of chemicals, and microorganisms including pathogens and multi-resistant bacteria from several sources which may be released into the environment ²⁴. The outcomes of this preliminary short study indicates that the wastewater samples from local sewage discharge can provide a rich source of L-asparaginase producing bacteria. The outcomes of this preliminary screening encourages to further investigate the sewage water samples to screen out the possibly new & efficient asparaginase producing isolates with complete biochemical & molecular characterization in view of commercial prospects. However, the bacterial isolates culture in this study will be optimized for enzyme production as well as purification.

REFERENCES

- 1. Karpel-Massler G, Ramani D, Shu C, Halatsch ME, Westhoff MA, Bruce JN, Canoll P and Siegelin MD. Metabolic Reprogramming of Glioblastoma Cells by L-asparaginase Sensitizes for Apoptosis *in vitro* and *in vivo*. Oncotarget. 2016; 7: 33512–33528.
- 2. Ghasemi A, Asad S, Kabiri M and Dabirmanesh B. Cloning and Characterization of Halomonas Elongata L-asparaginase, A Promising Chemotherapeutic Agent. Applied Microbiology and Biotechnology. 2017; 101: 7227–7238.
- 3. Radha R, Arumugam N and Gummadi SN. Glutaminase Free L-asparaginase from *Vibrio cholerae*: Heterologous Expression, Purification and Biochemical Characterization. International Journal of Biological Macromolecules; 2018; 111: 129–138.
- 4. Martinez-Outschoorn UE, Peiris-Pagés M, Pestell RG, Sotgia F and Lisanti MP. Cancer metabolism: a therapeutic perspective. Nature Reviews Clinical Oncology. 2017; 14: 11–31.
- 5. Laatsch H. Marine Bacterial Metabolites. In: Frontiers in Marine Biotechnology, Proksch, P. and Muller, W.E.G., eds. Horizon Bioscience, Norfolk, U.K. 2006; pp. 225–288.
- 6. Lebar MD, Heimbegner JL and Baker BJ. Cold-Water Marine Natural Products. Natural Product Report. 2007; 24: 774–797.
- 7. Anbu P, Gopinath SCB, Cihan AC and Chaulagain BP. Microbial Enzymes and Their Applications in Industries and Medicine. *BioMed Research International*, 2013; 204014.
- 8. Gopinath, SCB, Anbu P, Lakshmipriya T and Hilda A. Strategies to Characterize Fungal Lipases for Applications in Medicine and Dairy Industry. BioMed Research International. Article ID 154549, 10 pages, 2013.
- 9. Cammack KA, Marlborough DI and Miller DS. Physical Properties and Subunit Structure of Lasparaginase Isolated from *Erwinia carotovora*. Journal of Biochemistry. 1972;126: 361-379.
- 10. Manna S, Sinaha A, Sadhukhan R and Chakrabarty SL. Purification, Characterization and Antitumor Activity of L-asparaginase Isolated from *Pseudomonas stutzeri*. MB-405. Current Microbiology. 1995; 730: 291-298.

- 11. Abdel-Fatteh Y and Olama ZA. L-Asparaginase Produced by *Pseudomonas aeruginosa* in Solid State Culture: Evaluation and Optimization of Culture Conditions Using Factorial Designs. Process Biochemistry. 2002; 38: 115-122.
- 12. Qin M and Zhao F. L-asparaginase Release from *Escherichia coli* cells with Aqueous Two-phase Micellar Systems. Applied Biochemistry and Biotechnology. 2003;110(1):11-21.
- 13. Alrumman SA, Mostafa YS, Al-izran KA, Alfaifi MY, Taha TH and Elbehairi SE. Production and Anticancer Activity of an L-Asparaginase from *Bacillus licheniformis* Isolated from the Red Sea, Saudi Arabia. Scientific Reports. 2019; 9: 3756.
- 14. Sardessa YN and Bhosle S. Industrial Potential of Organic Solvent Tolerant Bacteria. Biotechnology Progress. 2004; 20(3): 655–660.
- 15. El-Naggar N, Moawad H, El-Shweihy NM and El-Ewasy SM. Optimization of Culture Conditions for Production of the Anti-Leukemic Glutaminase Free L-Asparaginase by Newly Isolated Streptomyces olivaceus NEAE-119 Using Response Surface Methodology. BioMed research international. 2015; 627031. https://doi.org/10.1155/2015/627031
- 16. Fatima N, Khan MM and Khan IA. L-Asparaginase Produced from Soil Isolates of *Pseudomonas aeruginosa* Shows Potent Anti-Cancer Activity on HeLa cells. Saudi Journal of Biological Sciences. 2019; 26: 1146–1153. https://doi.org/10.1016/j.sjbs.2019.05.001
- 17. Tomar RS, Sharma N and Kaushik S. Isolation and Characterization of L-Asparaginase Extracellular Enzyme Producing Bacteria from Industrial Soil Samples. International Journal of Pharmaceutical Sciences and Research. 2019; 10(11): 4937-4941.
- 18. Bhat MR, Nair JS and Marar T. Isolation and Identification of L-asparaginase Producing Salinicoccus sp. MKJ 997975 from Soil Microbial Flora. International Journal of Pharmaceutical Science and Research. 2015;6(8): 3599-05.
- 19. Talluri VSSLP, Bhavana M and Rajagopal SV. Isolation and Screening of L-Asparaginase Producing Bacteria from Visakhapatnam Soil Samples. International Journal of Pharmaceutical, Chemical and Biological Sciences. 2013; 3(4): 1121-1125.
- 20. Cyprowski M, Stobnicka-Kupiec A, Ławniczek-Wałczyk A, Bakal-Kijek A, Gołofit-Szymczak M and Górny RL. Anaerobic Bacteria in Wastewater Treatment Plant. International Archives of Occupational and Environmental Health. 2018; *91*(5), 571–579.
- 21. Cai L and Zhang T. Detecting human bacterial pathogens in wastewater treatment plants by a high-throughput shotgun sequencing technique. Environmental Science & Technology. 2013; 47: 5433–5441.
- 22. Plumb JJ, Bell J and Stuckey DC. Microbial Populations Associated with Treatment of an Industrial Dye Effluent in an Anaerobic Baffled Reactor. Applied and Environmental Microbiology. 2001;67(7): 3226-3235.
- 23. Liu Y, Dong Q and Shi H. Distribution and Population Structure Characteristics of Microorganisms in Urban Sewage System. Applied Microbiology and Biotechnology. 2015; 99(18): 7723–7734.
- 24. Numberger D, Ganzert L, Zoccarato L, Mühldorfer K, Sauer S, Grossart HP and Greenwood HP. Characterization of Bacterial Communities in Wastewater with Enhanced Taxonomic Resolution by Full-Length 16S rRNA Sequencing. *Science Reports*. 2019;9: 9673.