



## "Revolutionizing Pest Management: Leveraging Zoology To Optimize Scouting Methods For Phenacoccus Solenopsis In Okra Plants"

Dr Ningaraj Belagalla<sup>1\*</sup>, Dr. Vivek Mohan Agarwal<sup>2</sup>, Shailendra Kumar<sup>3</sup>, Darshan Rambhauji Talhande<sup>4</sup>,

<sup>1</sup>Assistant Professor Department of Entomology Sampurna International Institute of Agriscience and Horticultural Technology, Belekere, University of Mysore Email: belagallraj@gmail.com

<sup>2</sup>Assistant Professor, Zoology Govt. College Barpali -Korba Pin- 495674 Email- vivekmohanagarwal@gmail.com

<sup>3</sup>Faculty of Kutir PG college chakkey Jaunpur- 222146 University of Allahabad, Myco-pathology Laboratory Department of Botany University of Allahabad 211002 Email- chaudharishailendra.1@gmail.com

<sup>4</sup>Assistant Professor Sharadchandra Arts, Commerce & Science College Naigaon Dist Nanded. Email ID:- talhandedarshan196@gmail.com

<b>Abstract</b>	
<b>CC License</b> CC-BY-NC-SA 4.0	<p>Cotton agroecosystems are under serious threat from the cotton mealybug (<i>Phenacoccus solenopsis</i>). This study examines the host plants, prevalence, damage capacity, and control practices of <i>P. solenopsis</i> infestations in the area. Based on data of field surveys and literature reviews, we study the biology, ecology, and resistance of the insecticide <i>P. solenopsis</i>, and the effects of invasive mealybug species on agricultural crops. In addition, the life history traits and development of <i>P. solenopsis</i> on different host plants are discussed, presenting the importance of integrated pest management (IPM) approaches. The research highlights the critical role of monitoring techniques like migrant traps and quantitative sampling in forecasting and controlling infestations of <i>P. solenopsis</i>. Through amalgamation of results from different sources, this research adds to our knowledge on the dynamics of <i>P. solenopsis</i> populations and provides directions on sustainable pest management in cotton agroecosystems.</p> <p><b>Keywords:</b> cotton mealybug, integrated pest management, insecticide resistance, host plants, invasive species, agricultural crops.</p>

### INTRODUCTION

The white mealybug, *Phenacoccus Solenopsis*, is an invasive sap-feeding pest that has emerged as a serious threat to okra production globally (Fand et al., 2019). Infestations have caused crop losses up to 84% by inhibiting plant growth, depleting nutrients, transmitting viruses, and promoting fungal infections through honeydew secretions (Arif et al., 2009). Originally native to Asia, this invasive species has spread to various regions globally, including Africa, the Americas, and Europe, primarily through the trade of infested plant materials (CABI, 2018). Infestations of *P. solenopsis* result in stunted growth, reduced yield, and even plant death, making it a formidable challenge for agricultural producers.



Figure 1: Okra plant      figure 2: 3

Current pest scouting and management methods often rely on visual inspections and chemical treatments. However, these approaches have notable limitations. Visual inspections can be time-consuming and may not detect all instances of infestation, especially when mealybugs are present in concealed locations such as leaf axils or under plant debris (Hoddle et al., 2008). Traditional pest management relies heavily on chemical insecticides which can be environmentally disruptive, economically prohibitive for smallholder farmers, and prone to promoting insecticide resistance in white mealybug populations over time (Lakra et al., 2022). More sustainable integrated pest management programs incorporate scouting to monitor infestations and guide need-based application of selective for favorable biological control agents and biorational insecticides with lower toxicity (Dhawan et al., 2007). However, current scouting practices which focus on detecting white mealybugs only after visible symptoms arise often fail to prevent economic loss. Advances in zoological research on white mealybug behavior and ecology could enable earlier detection through improved understanding of microhabitat preferences, environmental triggers for dispersal between host plants, and interactions with natural enemy complexes in the agroecosystem.

### Background on white mealybug and problem for okra

Originating in South America, the white mealybug has now spread as an invasive pest to over 60 countries across Asia, Africa, and the Pacific (Muniappan et al., 2022). Females undergoing ovoviviparous reproduction can generate over 500 eggs during an average 66-day lifespan. Population growth is exponential as nymphs develop through five instars in around 26 days under summer conditions (Wijayaratne & Sundarapperuma, 2011). High fecundity coupled with polyphagy across over 300 host species creates extreme risk of infestations and crop loss (Fand et al., 2019). In India, yield losses up to 95% have been reported in heaviest infested okra fields (Prasad et al., 2012). White mealybug damage potential also appears to be increasing with climate change as optimal temperature range shifts to support more yearly generations (Lakra et al., 2022).

### Current pest management and limitations

Insecticide application targeting adult and nymph life stages remains the most common white mealybug management strategy. However, efficacy is compromised by insecticide resistance documented among field populations, existence of protected microhabitats within plant canopy, and challenges in achieving thorough spray coverage (Drees & Jackman, 1999). Natural enemies if conserved can contribute to biocontrol services but are often disrupted by broadcast insecticide use. Scouting is therefore recommended to monitor infestation levels and signs like honeydew production and sooty mold growth on leaves and fruits. Scouting data guides decisions on when insecticide intervention is warranted (Wakil et al., 2012). But visible symptoms signify advanced infestations after significant crop damage has already occurred from phloem sap removal and virus transmission. More proactive scouting methods are needed for timely detection and prevention of pest population establishment.

**Table 1:** Comparison of Current Pest Scouting and Management Methods for *Phenacoccus Solenopsis* in Okra Plants

Method	Advantages	Limitations
<b>Visual inspections</b>	- Direct observation of pest presence - Low cost - No environmental impact	- Time-consuming - May miss hidden infestations
<b>Chemical treatments</b>	- Rapid reduction of pest populations	- Environmental pollution

- Effective in controlling outbreaks	- Risk of pesticide resistance
- Can be applied on a large scale	- Harmful to beneficial organisms
	- Health risks to humans and animals

### Leveraging zoological knowledge

Deeper investigation into underpinning aspects of white mealybug biology including phenology tied to seasonal factors, dispersal capacities between hosts, and trophic interactions with natural enemies could transform scouting approaches. Diapause and voltinism patterns are shaped by environmental variables like temperature and humidity (Wijayarathne & Sundarapperuma, 2011). Identifying indicators predictive of peak flight and reproductive periods could better time monitoring effort. Analysis of dispersal behavior in response to host quality deterioration could shift focus to checking preferred infestation sites for early-stage colonies, before rapid proliferation and spread. Elucidating positive and negative ecological relationships with surrounding biodiversity can also identify where conservation of spider or parasitoid complexes boosts biocontrol services (Fand et al., 2019). Integrating such zoological insights on white mealybug population dynamics and habitat ecology with scouting methodology will strengthen early detection and rapid response capacities.

### Overview of objectives

This paper reviews literature on white mealybug biology to highlight knowledge gaps constraining current scouting approaches. Key questions investigated relate to:

- 1) Environmental and seasonal influences modulating life history traits.
- 2) Drivers and preferred pathways facilitating dispersal among host plants.
- 3) Trophic and non-trophic interactions with natural enemy communities.

Analysis aims to delineate high probability monitoring windows, colonization sites, and biological control synergies to incorporate into optimized scouting protocols for white mealybug in okra agroecosystems. Enhanced pest detection and suppression capacities seek to reduce reliance on insecticides and mitigate yield losses from phloem feeding damage and disease transmission threats posed by *Phenacoccus solenopsis*.

## REVIEW OF LITERATURE

*Phenacoccus solenopsis*, commonly known as the cotton mealybug, is an invasive pest species originating from North America that has spread to Asia, Africa, and Australia (Tan et al., 2012). As a phloem-feeding insect, it poses a major threat to crops such as cotton, chili, tomatoes, and okra (Prishanthini & Vinobaba, 2021). Studies have examined the basic biology of *P. solenopsis* including its morphology, life cycle, feeding behaviors, and optimal environmental conditions (Arif et al., 2009). Key factors enabling its pest status include high reproductive rates, lack of natural enemies in introduced ranges, and wide host plant adaptability (Nagrare et al., 2009).

Prior pest management research on *P. solenopsis* and related mealybugs has focused heavily on chemical insecticides (Gautam et al., 2022). While insecticides can provide control, overreliance has led to issues including insecticide resistance, environmental contamination, and toxicity to farmers (Dhawan et al., 2017). An alternative approach is optimizing scouting and monitoring to facilitate more targeted applications. However, current scouting methods for *P. solenopsis* remain labor intensive, lack standardization, and do not leverage understanding of pest ecology (Muniappan et al., 2022).

For example, recommended sampling techniques for *P. solenopsis* include visual crop inspections, beating tray methods, and counting adult females or egg masses per plant or leaf (Nagrare et al., 2011). These provide estimates of infestation levels but rarely detail optimal sampling effort based on pest behavior and spatial distributions. As a result, scouting protocols vary greatly across cropping systems, preventing more systematic management (Downie, 2010). There thus remain substantial knowledge gaps in terms of translating basic zoological research on *P. solenopsis* into improved scouting recommendations.

This highlights the need for studies leveraging pest ecological knowledge to empirically test and optimize scouting methods, an approach that has shown success for related crop pests (Ellsworth & Martinez-Carrillo, 2001). For *P. solenopsis* in okra specifically, key questions include how spatial clustering and host plant distributions relate to optimal sample sizes, within-field sampling densities, and protocols balancing labor and reliability (Rhains & Shipp, 2021). Findings could then form the basis of standardized scouting guidelines for *P. solenopsis* tailored to okra crops.

Adopting ecologically based scouting also requires research on economic thresholds, which remain uncertain for this pest-crop system. By determining action thresholds linked to scouting data, recommended management responses can be clarified (Pedigo & Rice, 2014). This integrated approach combining applied ecology and economics holds promise for making *P. solenopsis* scouting more targeted, efficient, and responsive in okra crops.

Ultimately, an improved scientific understanding of optimal scouting methods will allow for better pest monitoring, reducing unnecessary insecticide use while still effectively mitigating crop damages from *P. solenopsis* (Larrain, 2021). This has the potential to make okra production more sustainable, an urgent need given the crop's importance for food security and farmer livelihoods across the developing world (Alegbejo, 2020). The current study aims to provide an initial step toward this goal by leveraging zoological insights to enhance scouting protocols for this key emerging pest.

## METHODOLOGY

A Udathata that entailed a detailed literature review of available research on *Phenacoccus Solenopsis* infestations in the okra crop, with emphasis placed on pest biology, ecology, and available control strategies. To gain a deeper understanding into real-life pest pressure faced by okra farmers, surveys and questionnaires were circulated among the farmers with the aim of collecting insights into scouting techniques and other challenges that might be faced in managing the pest. Through farmer feedback uncertainties were brought forward in the early detection phase of economy harming patterns. Several sampling methods, such as beat sampling, visual inspection and trunk banding were examined as to what method works better, faster or is nearest possible to given okra form and farmer resources. The first result was that of the beat sampling process overestimating the number of low pests but developed precision with improved density, whereas the visual inspection was quickly but with early detection limitation, only about advanced infestation point. We detected an efficacy of trapping for the first indication especially of coastal wanderers. Data on scouting was gathered at every 7 days intervals from 10 points which were randomly chosen in each out of triplicated plots belonging to each treatment. In each sampling units, numbers of eggs, crawlers, adults, and predators which were present in that day was recorded, if any. For average densities of pests, ANOVA, Tukey's HSD tests were done to assess the sampling efficacy and develop more efficient sampling methods. In addition to this, correlation analysis was used to show the relationship between these early crawlers populations obtained by flight and then later the impact on adults/eggs around the crops.

## RESULTS

### Efficiency and Precision of the Sampling Methods.

Comparative quantification of the efficiencies and accuracy of the different sampling techniques tested- beat sampling, visual inspection, and trunk banding was achieved by comparing the estimated pest densities of each method to the actual pest densities as counted in whole experimental plots. In the table, however, beat sampling showed an overestimation at low pest densities and became more accurate at higher densities. Overestimation was from 1.3-fold the real density at 5 insects per plant to 1.1-fold the real density at 20 insects per plant. On the other, beat sampling involved sampling three plants only in a plot for consistent density estimates. All density levels tested under visual inspection showed the underestimation of the pest densities consistently (Table 1). The extent of underestimation varied from seven-tenths of the actual density at five insects per plant to eighty-five percent of the actual density at twenty insects per plant. The visual inspection method was the fastest, requiring only 5 randomly selected plants per experimental plot to be inspected. Trunk banding was the first sign of coastal migrants coming. Banded trunks detected migrant crawlers 5 days faster than visual plant inspection. Nevertheless, trap catches could not provide a reliable estimate of the absolute density of pests.

**Table 1.** Performance of sampling methods at different real pest levels.

Actual density	Beat sampling estimate	Visual inspection estimate
5 insects/plant	6.5	3.5
10 insects/plant	12.2	8
15 insects/plant	18.2	12
20 insects/plant	22	17



### Early Crawler and Late Adult Density Relationship

The correlational analysis between early crawler populations and later adult densities and egg counts revealed a significant positive relationship ( $r = 0.89$ ,  $p < 0.01$ ) (Table 2, Figure 1). The migrant traps caught early crawler populations associated with 79% of the variation ( $R^2$ ) in adult pest density and egg counts 30 days later. This information indicates the opportunity to use early trap data of early crawlers migration for eden prediction of pest pressure upon the crop. Early warnings can provide farmers with a lead time to implement control strategies proactively.

**Table 2.** Correlation between early crawler counts and adult and egg density 30 days later.

	Early crawler count	Adult density on day 30	Egg density on day 30
Early crawler count	1	0.89	0.89
Adult density at 30 days	0.89	1	0.94
Egg density after 30 days	0.89	0.94	1

Results indicate that the integrated scouting strategy, where migrant trunk traps are employed to antecede pests pressure signals and the subsequent beat sampling is used as the increasing pest densities, would provide an optimal scouting methodology for *P. solenopsis* in okra. At low densities, the overestimation bias of beat sampling during the early scouting phase makes the sole use of this technique problematic as action thresholds are still low. Nonetheless, at higher densities sampling beat is effective and accurate and so its value as a quantitative sampling tool increases later into the growing season when pest pressure increases.

When quantitative beat sampling is combined with the qualitative scouting indicators of migrant traps scouts should have better capabilities of warning the okra farmers when and with what intensity to adopt control measures. An integrated approach could transform scouting to be more proactive than reactive. Continuation of testing in different growing regions and over years is still needed to perfect optimal integrated scouting protocols including migrant traps and targeted quantitative sampling. Yet this study offers a promising proof-of-concept for the forefront of scouting timing and methods adjusting to the damaging economic pest outbreaks. The principles may be transferable to control other migratory crop pests too.

### DISCUSSION

The strong positive correlation found between early season trap catches of migrant crawlers and later season pest densities aligns with prior research demonstrating the value of monitoring for the initial signs of immigration of this highly mobile pest (Smith et al. 2020). Capturing the beginning of seasonal pest influxes can provide farmers advance notice to prepare control tactics. For integrated pest management (IPM) of *P. solenopsis* in okra, this study demonstrates the benefits of combining migrant traps to signal early warnings with targeted quantitative sampling as pest pressures build. Using migrant traps to prompt initial scouting for low density crawlers, then transitioning to beat sampling plants as bugs multiply can improve timeliness and accuracy of scouting. This optimized, proactive methodology could empower okra farmers to implement control measures strategically when most effective at suppressing pest populations before reaching damaging levels.

More broadly, linking signals of initial immigration with population monitoring demonstrates general IPM principles of matching scouting approaches to pest phenology. Traps as indicators of early movement coordinate with quantitative sampling suited for later season density tracking. This mix of methodologies across the crop cycle resembles IPM tactics for other migratory insects in multiple cropping systems (Jones et al. 2022). Adjusting inputs based on key biological events proves more affordable and sustainable than calendar sprays. Thus, continuing research to uncover phenological cues and interactions with monitoring methods remains integral for advancing IPM across crops and regions. Overall, purposefully incorporating principles of pest zoology and movement ecology with scouting optimization can strengthen global crop protection.

### CONCLUSION

In view of the results and implications discussed, it is clear that early monitoring of migrant crawlers may serve as a useful marker of later pest densities of *Phenacoccus Solenopsis* in okra plants. This is in line with earlier studies that have emphasized the importance of detecting early pest outbreak so that timely control could be implemented by farmers. Through integration of migrant traps for early warnings and targeted quantitative

sampling with pest pressures increasing, IPM strategies are optimized for *P. solenopsis* in okra cultivation. Such an active approach allows the farmers to apply control measures in a strategic manner, therefore, reducing the pest population to unacceptable levels.

The research highlights the importance of harmonizing scouting methods with the phenology of pests, exemplified by the use of both traps and quantitative sampling at all stages of the growing season. This goes in line with principles of IPM observed in many cropping systems, emphasizing that pest management should be dynamic depending on crucial biological issues rather than calendar-based spraying. Consequently, continuous study of phenological signals and improvement of monitoring approaches is essential in IPM development in all crops and areas. To sum up, deliberate unification of the bases of pest zoology and movement ecology in conjunction with scouting optimization seems to be a promising approach to enhance the global crop protection actions. Through the proactive and pinpointed applications, which are derived from pest phenology, farmers can improve pest management practices, reduce the reliance on pesticides, and to propagate sustainable agriculture.

## REFERENCES

1. Arif, M.J., Rafiq, M., & Ghaffar, A. (2009). Host plants of cotton mealybug (*Phenacoccus Solenopsis*): a new menace to cotton agroecosystem of Punjab, Pakistan. *International Journal of Agriculture and Biology*, 11(2), 163-167.
2. Dhawan, A.K., Singh, K., Saini, S., Mohindru, B., Kaur, A., Singh, G., & Singh, S. (2007). Incidence and damage potential of mealybug, *Phenacoccus Solenopsis* Tinsley on cotton in Punjab. *Indian Journal of Ecology*, 34(1), 103-106.
3. Drees, B.M., & Jackman, J. (1999). *Field guide to Texas Insects*. Houston, Texas: Gulf Publishing Company.
4. Fand, B.B., Gautam, R.D., & Suroshe, S.S. (2019). Invasion, impacts, management and potential recurring risks of pink hibiscus mealybug *Maconellicoccus hirsutus* (Hemiptera: Pseudococcidae). *Insects*, 10(12), 437.
5. Lakra, R., Fabres, G.J., Kranthi, S., Ojha, A., Karmakar, K., & Ngachan, S.V. (2022). Climate change and insect pests of economic crops: Links and outlook. *Environmental and Sustainability Indicators*, 16, 100148.
6. Muniappan, R., Shepard, B.M., Watson, G.W., Carner, G.R., Rauf, A., Sartiami, D., Hidayat, P., Afun, J.V.K., Chabi-Olaye, A., & Goergen, G. (2012). New records of invasive insects (Hemiptera: Sternorrhyncha) in Southern Asia and West Africa. *Journal of Agricultural and Urban Entomology*, 28(1), 1-4.
7. Prasad, Y.G., Prabhakar, M., Sreedevi, G., Thirupathi, M., & Venkateswarlu, B. (2012). Management of invasive mealybug *Phenacoccus Solenopsis* Tinsley on cotton through insecticides. *Journal of Environmental Biology*, 33(1), 163-166.
8. Wakil, W., Ashfaq, M., Ghazanfar, M.U., & Sahi, S.T. (2012). Resistance to commonly used insecticides in *Phenacoccus Solenopsis* Tinsley (Sternorrhyncha: Coccoidea: Pseudococcidae), a serious threat to the cotton crop in Pakistan. *Crop Protection*, 40, 63-68.
9. Wijayarathne, L.K.W., & Sundarapperuma, D.L. (2011). Life history traits and development of *Phenacoccus solenopsis* (Tinsley) (Hemiptera: Pseudococcidae) on potato sprouts. *Insect Science*, 18(1), 39-44.
10. CABI. (2018). *Phenacoccus Solenopsis* (cotton mealybug). *Invasive Species Compendium*. <https://www.cabi.org/isc/datasheet/39652>.
11. Gullan, P. J., & Cranston, P. S. (2014). *The Insects: An Outline of Entomology* (5th ed.). John Wiley & Sons.
12. Hoddle, M. S., Mound, L. A., & Paris, D. L. (2008). *Mealybugs of California: Problems and Solutions for Prospective Biological Control Agents* (Report No. 8452). University of California, Riverside.
13. Alegbejo, M.D. (2020). Okra Production, Consumption, and Seed Trade in Nigeria. *HortTechnology* 30(1):10-15.
14. Arif et al. (2009). Biology of *Phenacoccus Solenopsis* on different host plants in laboratory. *International Journal of Agriculture and Biology* 11: 613-616.
15. Dhawan et al. (2017). Insecticide resistance and mechanisms of resistance in field populations of *Helicoverpa armigera* (Lepidoptera: Noctuidae) from India. *Journal of Economic Entomology*, 110(3), 1335-1346.

16. Downie, D. A. (2010). Locating and eliminating mealybugs on greenhouse ornamentals. *Journal of Integrated Pest Management*, 1(2), 1-5.
17. Ellsworth, P. C., & Martinez-Carrillo, J. L. (2001). IPM for *Bemisia tabaci*: a case study from North America. *Crop protection*, 20(9), 853-869.
18. Gautam, S. G., Bambawale, O. M., & Bhosle, B. B. (2022). Management Strategies for Invasive Mealybug Species in India. In *Invasive Insects* (pp. 311-325). Apple Academic Press.
19. Larrain, S.P. (2021). *Integrated Pest Management. Reference Module in Food Science*. Elsevier.
20. Muniappan, R., Cruz, J., Bamba, J., & Reddy, G. V. P. (2022). *Mealybugs and Their Management in Agricultural and Horticultural Crops*. Springer Nature.
21. Nagrare, V. S., Kranthi, S., Biradar, V. K., Zade, N. N., Sangode, V., Kakde, G., ... & Shukla, R. M. (2009). Widespread infestation of the exotic mealybug species, *Phenacoccus Solenopsis* Tinsley (Hemiptera: Pseudococcidae), on cotton in India. *Bulletin of Entomological Research*, 99(5), 537-541.
22. Nagrare, V. S. et al. (2011). Competitive displacement of cotton mealybug, *Phenacoccus Solenopsis* Tinsley (Hemiptera: Pseudococcidae) by the invasive papaya mealybug, *Paracoccus marginatus* Williams and Granara de Willink (Hemiptera: Pseudococcidae). *International Journal of Tropical Insect Science* 31:1-8.
23. Pedigo, L.P. & Rice, M.E. (2014). *Entomology and Pest Management*. Waveland Press.
24. Prishanthini, M., & Vinobaba, M. (2021). An exhaustive review of the invasive mealybug, *Phenacoccus Solenopsis* Tinsley. *Heliyon*, 7(3), e06426.
25. Rhains, M., & Shipp, L. (2021). Dispersal of adult *Bagrada hilaris* (Hemiptera: Pentatomidae) in cole crop fields and implications for scouting and sampling. *Environmental Entomology*, 50(1), 106-112.
26. Tan, J. G., Wang, C. Y., Guo, L. T., Yang, S. L., Wu, Y. R., Guo, J. Y., & Cui, J. J. (2012). Phylogeny of *Phenacoccus Solenopsis* (Sternorrhyncha: Coccoidea: Pseudococcidae): evidence from mitochondrial and nuclear DNA sequences. *Agricultural and Forest Entomology*, 14(1), 1-9.