



Isolation, Characterization and Screening of Microbial Melanin for its role in Protection of Plant Growth Promoting Bacteria

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Abstract

Plants are susceptible to vulnerable impacts caused by presence of heavy metals, viruses, bacteria, fungi, insects and pests. Along with diseases, availability of nutrients, water, temperature and the role of protecting the role plant growth promoting bacteria (PGPB) is also a matter of concern. PGPB may promote plant growth directly usually by either facilitating resource acquisition or modulating plant hormone levels, or indirectly by decreasing the inhibitory effects of various heavy metals on plant growth and development. Although a significant increase in the use of PGPB in agriculture was observed in the last two decades, there is a dearth of long-term studies addressing the effects of PGPB on existing microbial community structure. Melanin is a negative charge hydrophobic complex pigment that is a substance made of small particles virtually insoluble in the environment and is usually used for its color, protective or other characteristics. Melanin, its ability to chelate metals by which the toxicity of metals can be reduced. In this direction, soil and vegetable waste samples were collected and enriched. A total of fourteen isolates were obtained and these isolates were screened for melanin production. Black brownish colonies were melanin positive colonies. These isolates were subjected to production of melanin. Melanin was extracted and subjected to FTIR (Fourier Transform Infrared Spectroscopy). On the basis of FTIR results, melanin was confirmed and then tested for its metal chelating ability followed by its application as protectant for plant growth promotion. Significant growth was observed when compared with control proves its vital role in plant growth promotion.

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Keywords: PGPB, Melanin, FTIR, Growth promotion

Introduction

Melanin is an ancient pigment that occurred very early in all living organisms. Melanin is typically known for its unique ability to absorb a wide range of radiations ⁵. Melanin is a bioactive-complex pigment which helps to protect the microorganisms from environmental stresses. Pigmented compounds from microbes are provisionally classified as melanin on the basis of their physical and chemical properties ⁶.

Melanin pigments were extracted from a wide variety of microorganisms including bacteria and fungi. In the present study isolation, identification and characterization of melanin ⁷. Melanin is a common name for a group

of biopolymers with the dominance of potential applications in medical sciences, cosmeceutical, bioremediation, and bioelectronic applications⁸. Actinomycetes are characterized by the production of various pigments on natural or synthetic media⁹. Actinomycetes often show themselves to have outstanding antimicrobial activity¹⁰. Actinomycete are biotechnologically important microorganism and they produce many pigments with industrial applications is a diffusible, dark pigment which is water soluble.⁷ Actinomycetes also synthesizes and excrete dark pigments, melanin or melanoid, which are considered to be a useful criterion for taxonomic studies.⁸

Melanin compounds are irregular, dark brown polymers that are produced by various microorganisms by the fermentative oxidation, and have the radioprotective and antioxidant properties that can effectively protect the living organisms from ultraviolet radiation¹¹. Melanin's are used as the adsorbents of radionuclides and heavy metal ions¹², show strong biological activities in the clearance of free radicals and protection against radiation¹³, improve the survival rate under serious environmental stressors, such as extreme temperatures, drought, irradiation and metals exposure¹⁴, and can substitute for synthetic pigments¹⁵. These pigments are usually described in terms of various shades of blue, violet, red rose, yellow, green, brown and black. The pigments may be dissolved into the medium or it may be retained in the mycelium¹⁶. HPLC is used for quantification of melanin, particularly when it is necessary to distinguish between eumelanin and pheomelanin¹⁷.

Actinomycetes are known to produce various kinds of antibiotics and moreover these antibiotics include many pigments. Production of pigments by actinomycetes has been utilized as an important cultural characteristic in describing the organisms⁹. Melanogenesis in nature is initiated through the activation of tyrosinase, laccase and other polyphenol oxidases in response to external stimuli such as chemical stresses (metal ions, ROS, oxidizing agents), UV exposure, and pathogenic attack (particularly in insects)¹⁸.

Actinomycetes are Gram-positive bacteria showing a filamentous growth. They are a group of organisms widespread in nature, and play a significant role in the future of biotechnology, because of their importance as producers of vitamins, enzymes, antitumor agents, immune modifying agents and mainly antibiotic compounds¹⁹.

The current work focuses on application of melanin for improvement of plant growth promotion.

Materials and Methods

Sample collection, enrichment of samples and isolation of microorganisms

Soil samples were collected from the local garden area, vegetable dumping area and Karnala region, Panvel, Navi Mumbai. One gram of the collected samples were inoculated into an enrichment medium namely sterile tyrosine casein broth. Enrichment broth was incubated at room temperature for 48 – 72 hours followed by isolation of microorganisms into sterile tyrosine casein agar plates. Plates were incubated at room temperature for 48 – 72 hours

Screening of melanin producing isolates

Melanin producing microorganisms were identified by the presence of microbial colonies with brown black color (diffusible) in agar plates. Selective colonies were separated out and streaked on tyrosine casein agar slants, sterile tyrosine casein agar plates for further characterization.

Production and extraction of melanin

Melanin production was carried out using sterile tyrosine casein broth by inoculating the isolates followed by incubation at room temperature for five days. Filtrate of tyrosine casein broth culture adjusted to pH-7. 0.5g of potassium persulphate was added to each of the filtrates. Allow to stand for two hours. at room temperature. After two hours, methanol was added to each filtrate. Allow to stand for three days at room temperature. Both filtrates were centrifuged at 10,000 rpm for 10 minutes. Pellet was dried and subjected to characterization by FTIR.

Characterization of melanin using Fourier Transform Infrared Spectroscopy

Sample was mixed with KBr powder and then compressed and then the sample was subjected to FTIR.

Application of melanin for plant growth promotion

Rice seeds (*Oryza sativa*) were mixed with extracted melanin and growth parameters like seed germination percentage, root length and shoot height were measured.

Germination Percentage (GP) = $D/E \times 100\%$

D is the number of germinated seeds on the seventh day,

E is the number of total seeds investigated

Results and Discussion

Sample collection, enrichment of samples and isolation of microorganisms

Enriched sample was serially diluted followed by isolation on to Tyrosine agar. The black brownish colonies were observed. These colonies were selected and isolated on a tyrosine agar plate and were kept for 48 hours at room temperature. Melanin producing microorganisms was identified by the presence microbial colonies with brown black color (diffusible) in agar plates

Selective colonies were separated out and streaked on Tyrosine agar slants, sterile Tyrosine agar plates for further characterization. A total of fourteen isolates were obtained after 48 of incubation. The colonies were screened for black brownish coloration. Those colonies which are black brownish in nature were selected for further research. As everyone knows the pigment producing microbes were found abundant in soil and vegetable waste.

Maintenance of pigment producing organism:

It is essential to maintain the obtained isolates throughout the research. For their maintenance, tyrosine agar was used. Single colony was streaked on slant and incubated at room temperature for 48 hours and then slants were wrapped with paper to reduce the chances of contamination. Slants were stored at 10°C. After 20-25 days culture was re-streaked for maintaining the fresh culture.

Production and extraction of melanin:

To characterize and determine the application of melanin, it was produced in bulk using the isolates and production media. Production was carried out in liquid media i.e., tyrosine broth. Optical density of 0.1 was adjusted using saline as blank and was inoculated into the production media. The broth was kept on shaker for 5 days and growth was monitored and also melanin production was checked after 5 days. Within 3 days a faint red color was observed in the inoculated broth. After 5 days the intensity of color increases and it changes to the darkish red and brown color and extraction of melanin was carried out as mentioned previously.

Characterization of melanin using Fourier-Transform Infrared spectroscopy

Peaks observed near 3300 cm^{-1} may be due to characteristic O-H stretching or N-H stretching vibrations of the carboxylic acid, and phenolic groups in melanin. Melanin form isolates K7, V1(8), V4(9) confirmed by FT-IR studies. The results of FTIR are shown in figure no. 1, 2, 3.

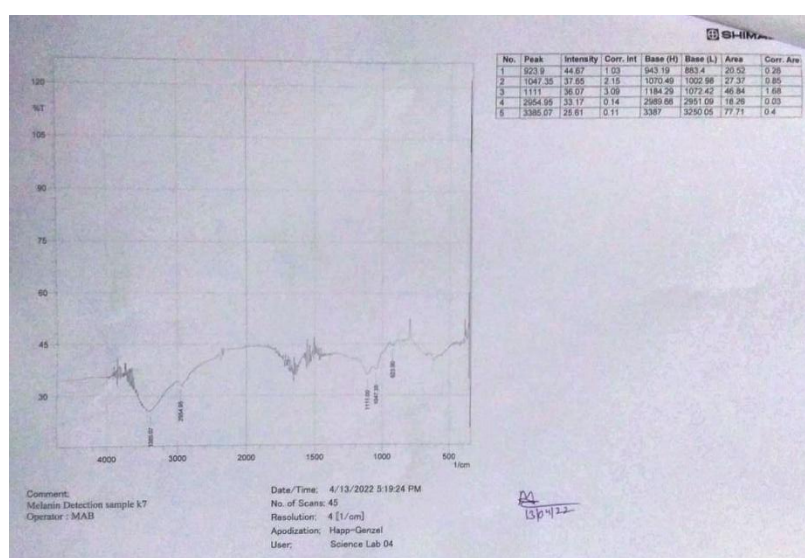


Figure No. 1: FTIR result of melanin from isolate K7

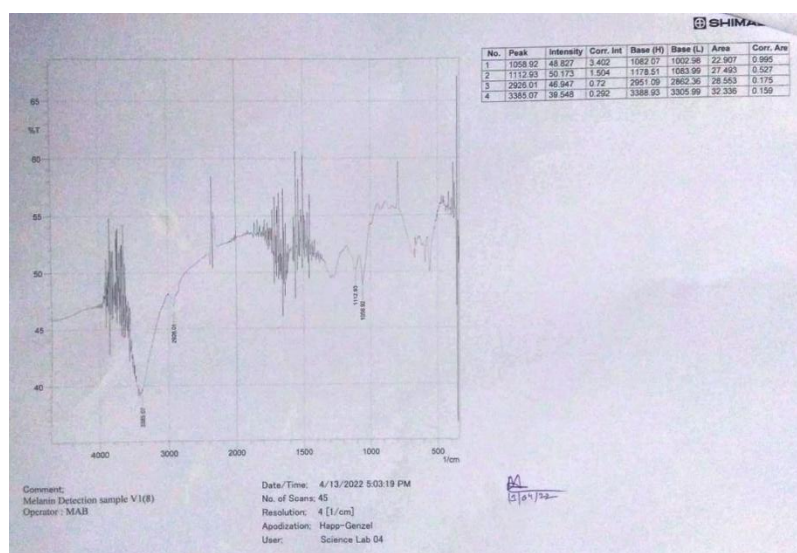


Figure No. 2: FTIR result of melanin from isolate V1(8)

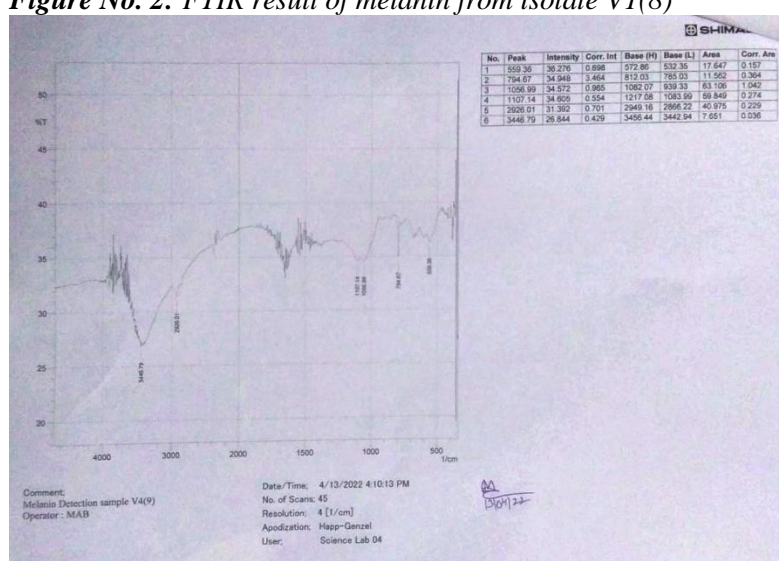


Figure No. 3: FTIR result of melanin from isolate V4(9)

Application of melanin for plant growth promotion

Seeds were mixed with melanin and germination percentage, root and shoot length were calculated. It was measured that melanin from isolate K7 was efficient in improving seed germination (82%) when compared with control (55%). Shoot length of treated plantlets showed 15.3cm while control showed 11.2cm and root length of treated plantlets measured as 13.1cm while control was 8.6cm. This indicates bacterial melanin helps in improvement of plant growth parameters.

Reference

1. Pralea IE, Moldovan RC, Petrache AM, et al. From Extraction to Advanced Analytical Methods: The Challenges of Melanin Analysis. *International Journal of Molecular Sciences*. 2019;20(16):3943. doi:10.3390/ijms20163943
2. Pralea IE, Moldovan RC, Petrache AM, et al. From Extraction to Advanced Analytical Methods: The Challenges of Melanin Analysis. *International Journal of Molecular Sciences*. 2019;20(16):3943. doi:10.3390/ijms20163943
3. Kimura S, Kurasaki M, Saito T, et al. Synthetic dopamine melanins, a model for neuromelanin, show superoxide dismutase-like activity. *Trace Elements and Electrolytes*. 2004;21(04):55-59. doi:10.5414/TEP21055
4. Pavan ME, López NI, Pettinari MJ. Melanin biosynthesis in bacteria, regulation and production perspectives. *Applied Microbiology and Biotechnology*. 2020;104(4):1357-1370. doi:10.1007/s00253-019-10245-y

5. Tran-Ly AN, Reyes C, Schwarze FWMR, Ribera J. Microbial production of melanin and its various applications. *World Journal of Microbiology and Biotechnology*. 2020;36(11). doi:10.1007/s11274-020-02941-z
6. Riddhi Naresh D, Madhava Anil K, Kalpana HM. Production and Characterization of Melanin from *Streptomyces Cavourensis* Strain RD8 Using Response Surface Optimization. *Environmental Pollution and Protection*. 2017;2(4). doi:10.22606/epp.2017.24002
7. Sivaperumal P, Kamala K, Rajaram R, Mishra SS. Melanin from marine *Streptomyces* sp. (MVCS13) with potential effect against ornamental fish pathogens of *Carassius auratus* (Linnaeus, 1758). *Biocatalysis and Agricultural Biotechnology*. 2014;3(4):134-141. doi:10.1016/j.bcab.2014.09.007
8. Singh S, Nimse SB, Mathew DE, et al. Microbial melanin: Recent advances in biosynthesis, extraction, characterization, and applications. *Biotechnology Advances*. 2021;53. doi:10.1016/j.biotechadv.2021.107773
9. Amal AM, Abeer KA, Samia HM, El-Nasser Nadia AH. Selection of Pigment (Melanin) Production in *Streptomyces* and Their Application in Printing and Dyeing of Wool Fabrics. Vol 1.; 2011. www.isca.in
10. Prakash U, Patnayak B. A Brief Review on: Production and Characterization of Antibiotic from *Streptomyces* Family Mini-Review. *Research and Reviews: Research Journal of Biology*. 3(2).
11. Dastager S, Li WJ, Tian XP, et al. Separation, identification and analysis of pigment (melanin) production in *Streptomyces* Fabrication of nanocomposites for inhibiting the ESBL producing bacteria and various cancer cells View project Phylogenomics of order Bifidobacteriales View project Separation, identification and analysis of pigment (melanin) production in *Streptomyces*. *African Journal of Biotechnology*. 2006;5(8):1131-1134. <http://www.academicjournals.org/AJB>
12. v. G. Babitskaya, v. V. Shcherba. The Nature of Melanin Pigments of Several Micro- and Macromycetes. 2002;38:247-251.
13. Gauslaa Y, Solhaug KA. Fungal melanins as a sun screen for symbiotic green algae in the lichen *Lobaria pulmonaria*. *Oecologia*. 2001;126(4):462-471. doi:10.1007/s004420000541
14. Mahendra Kumar C, Sathisha UV, Dharmesh S, Rao AGA, Singh SA. Interaction of sesamol (3,4-methylenedioxypheanol) with tyrosinase and its effect on melanin synthesis. *Biochimie*. 2011;93(3):562-569. doi:10.1016/j.biochi.2010.11.014
15. Li C, Ji C, Tang B. Purification, characterization and biological activity of melanin from *Streptomyces* sp. 1. *FEMS Microbiology Letters*. doi:10.1093/femsle/fny077/4975772
16. Fernandes B, Matamá T, Guimarães D, Gomes A, Cavaco-Paulo A. Fluorescent quantification of melanin. *Pigment Cell and Melanoma Research*. 2016;29(6):707-712. doi:10.1111/pcmr.12535
17. Park J, Moon H, Hong S. Recent advances in melanin-like nanomaterials in biomedical applications: A mini review. *Biomaterials Research*. 2019;23(1). doi:10.1186/s40824-019-0175-9
18. Vasanthabharathi V, Lakshminarayanan R, Jayalakshmi S. Melanin production from marine *Streptomyces*. *African Journal of Biotechnology*. 2011;10(54):11224-11234. doi:10.5897/ajb11.296
19. Plonka PM, Grabacka M. Melanin synthesis in microorganisms - Biotechnological and medical aspects. *Acta Biochimica Polonica*. 2006;53(3):429-443. doi:10.18388/abp.2006_3314
20. Tsai HF, Fujii I, Watanabe A, et al. Pentaketide Melanin Biosynthesis in *Aspergillus fumigatus* Requires Chain-length Shortening of a Heptaketide Precursor. *Journal of Biological Chemistry*. 2001;276(31):29292-29298. doi:10.1074/jbc.M101998200
21. Kwon-Young Choi 2. Bioprocess of Microbial Melanin Production and Isolation. Published online November 16, 2021.
22. Deshmukh KR. Isolation, Characterization of Melanin Producing Organism and Extraction of Melanin. *International Journal of Scientific & Engineering Research*. 2012;3(11). <http://www.ijser.org>
23. Guo J, Rao Z, Yang T, Man Z, Xu M, Zhang X. High-level production of melanin by a novel isolate of *Streptomyces kathirae*. *FEMS Microbiology Letters*. 2014;357(1):85-91. doi:10.1111/1574-6968.12497
24. Tomita K, Oda N, Ohbayashi M, Kamei H, Miyaki T, Oki T. A New Screening Method for Melanin Biosynthesis Inhibitors Using *Streptomyces bikiniensis*.
25. A.R.Srividya* RK and VJV. Isolation, identification, bioprocessing and characterization of secondary metabolites for its antimicrobial and genotoxicity from the soil screened microorganism. Published online November 2014.
26. Umesh Kumar*1 2, Brajesh Kumar 3, Anil Bhandari 4 and Y. Kumar 1. Phytochemical Investigation and Comparison of Antimicrobial Screening of Clove and Cardamom. Published online November 2010.
27. Madhusudhan DN, Mazhari BBZ, Dastager SG, Agsar D. Production and cytotoxicity of extracellular insoluble and droplets of soluble melanin by *Streptomyces lusitanus* DMZ-3. *BioMed Research International*. 2014;2014. doi:10.1155/2014/306895

28. Singh V, Haque S, Singh H, et al. Isolation, screening, and identification of novel isolates of actinomycetes from India for antimicrobial applications. *Frontiers in Microbiology*. 2016;7(DEC). doi:10.3389/fmicb.2016.01921
29. Vasanthabharathi V. Review on Melanin from Marine Actinomycetes. *Journal of Basic & Applied Sciences*. 2020;16(1):39-42. doi:10.29169/1927-5129.2020.16.05
30. Sambamurthy K, Ellaiah P. A new streptomycete producing neomycin (B&C) complex-S. marinensis (Part I). *Hindustan Antibiot Bull*. 17(1-2):24-28.
31. Noble K Kurian, Harisree P Nair, Sarita G Bhat. Melanin producing *Pseudomonas stutzeri* BTCZ10 from marine sediment at 96 m depth (Sagar Sampada cruise #305). 2014;2(5)(2321-8371):6-11.
32. Shoumita Chakrabarty SP. Isolation of Melanin Pigment Producing Bacteria from Marine Water and Study of Photoprotective Role of the Pigment. Published online September 2018.
33. D. N. Olennikov, S. V. Agafonova, A. V. Stolvikova, A. V. Rokhin. Melanin of *Laetiporus sulphureus* (Bull.: Fr.) Murr sterile form. 2011;47:298-303.
34. Guo J, Rao Z, Yang T, Man Z, Xu M, Zhang X. High-level production of melanin by a novel isolate of *Streptomyces kathirae*. *FEMS Microbiology Letters*. 2014;357(1):85-91. doi:10.1111/1574-6968.12497
35. Jacobson ES. Pathogenic Roles for Fungal Melanins. *Clinical Microbiology Reviews*. 2000;13(4):708-717. doi:10.1128/CMR.13.4.708
36. Glagoleva AY, Shoeva OY, Khlestkina EK. Melanin Pigment in Plants: Current Knowledge and Future Perspectives. *Frontiers in Plant Science*. 2020;11. doi:10.3389/fpls.2020.00770
37. S. Ceccarelli, S. Grando, J. A. G. Van Leur. Genetic diversity in barley landraces from Syria and Jordan. 1987;36:389-405.
38. Kovacs D, Flori E, Maresca V, et al. The Eumelanin Intermediate 5,6-Dihydroxyindole-2-Carboxylic Acid Is a Messenger in the Cross-Talk among Epidermal Cells. *Journal of Investigative Dermatology*. 2012;132(4):1196-1205. doi:10.1038/jid.2011.457
39. Kiran GS, Jackson SA, Priyadharsini S, Dobson ADW, Selvin J. Synthesis of Nm-PHB (nanomelanin-polyhydroxy butyrate) nanocomposite film and its protective effect against biofilm-forming multi drug resistant *Staphylococcus aureus*. *Scientific Reports*. 2017;7(1):9167. doi:10.1038/s41598-017-08816-y