

Introduction:

Organophosphorus pesticides are xenobiotic compounds that act as inhibitors of neuro transmitters such as acetylcholinesterase leading to neurological disorders and cancer related diseases when accumulated into food chain and food web. Soils have the capability of dissolving them but it takes longer periods (Satish et al., 2017). Annually millions of deaths and chronic poison in there were reported due to the residual deposition of these pesticides in soil, water as well as air (Frazer, 2000). The pesticides such as Chloropyrifos, parathion, methylparathion, glyphosate and Chlorantraniliprole are dangerous to mammals as their half-life in degradation in the soil takes much time (Singh and Walker, 2006) so, without disturbing the balance of ecological systems and one such alternative is using of microorganisms based degradation methods.

Microorganisms such as bacteria are able to metabolize these xenobiotic compounds into organic compounds through various degrading pathways. The enzymes produced by these organisms help in dissolution of the harmful active ingredients of pesticides such as Coumaphos degraded by *Falavobacterium* and *Achrobacterium* enzymes degrades Carbofuran (Mustapha et al., 2019). Organophosphorus pesticides are degraded by bacteria because those bacteria produce OPH enzyme which helps in hydrolysing of these pesticide class. In the present study an attempt was done to screen the bacteria that produce OPH enzyme which were of agriculture soil origin.

Experimental:

Soil sample collection and culturing of organisms:

Soil samples were collected from three study areas of Telangana region of Khammam district from tomato fields where agriculture practice was done from a decade so that microorganisms diversity can be found. The collected soil samples were serially diluted to reduce the microbial load in order to get isolated colonies. Then the last dilution factor that is 10^{-6} was taken and spread onto pre sterilized and prepared nutrient agar plates in sterile conditions and incubated at 37°C for 24 hours to get the microbial growth. After incubation, the growth was observed and loopful of colonies were inoculated streaking of the isolated colonies was done onto fresh nutrient agar plates which were sterile for sub-culturing.

Details of Chlorantraniliprole (Gade Pavan et al., 2023).

Common name	: Chlorantraniliprole
Molecular weight	: $483.15 \text{ g mol}^{-1}$
Colour	: White or colourless crystalline solid
IUPAC Formula	: 5-bromo-N-[4-chloro-2-methyl-6-(methylcarbamoyl)phenyl]-2-(3-chloropyridin-2-yl)pyrazole-3-carboxamide
Chemical Formula	: $\text{C}_{18}\text{H}_{14}\text{BrCl}_2\text{N}_5\text{O}_2$
Stability	: Stable in air and is not sensitive to UV radiation, it is stable to neutral and weakly acidic solutions, but is hydrolysed by strong bases.

Structure of Chlorantraniliprole:



Figure 1: Chlorantraniliprole structure (Aghris et al., 2022).

Optimization conditions followed for PDB isolation:

The accelerated degradation of pesticide Chlorantraniliprole whose structure is shown in figure 1, was found in minimal salt media at 37°C, pH7 in 5 hours 30 minutes when sucrose was added as substrate and pesticide was 10 mg/L, and the methodology was mentioned in (Gade Pavan et al., 2023).

Microbial growth on selective media:

The morphology of the colonies were observed under microscope such as size, shape, colour, spore formation and gram's stain, then selective media was prepared by using sucrose as one of the ingredient in nutrient agar medium composition as shown in table 1. Then the colonies were spread onto the selective media plates and pesticide of 10 mg concentration was used to further screen the colonies and the plates were incubated at 37°C for 24 hours.

Table 1: Composition of selective media (Minimal Salt media)

S.No	Minimal salt media ingredients	Composition
1.	K ₂ HPO ₄	0.6g/L
2.	KH ₂ PO ₄	0.2g/L
3.	MgSO ₄ .7H ₂ O	0.08g/L
4.	NaCl	0.2g/L
5.	NH ₄ NO ₃	0.6g/L
6.	Sucrose	2.0 g/L
7.	Distilled water	1000mL
8.	pH	7.0
9.	Agar	4grams

Screening of colonies showing pesticide degradation:

After incubation for 24 hours on selective media with pesticide Chlorantraniliprole, the zone of clearance were observed which were measured using a ruler.

Results and discussion:

The soil samples collected from Khammam district from the Tomato fields were serially diluted and the serial dilution concentration of 10⁻⁶ was spread onto sterile nutrient agar media plate and incubated. Then from the colonies formed on the plate a loopful of colonies were streaked onto fresh nutrient agar media to get isolated colonies as shown in figure 2.

**Figure 2: Isolated bacterial colonies****Morphological studies:**

The obtained isolated colonies were observed for morphological features which revealed that the organism was motile, rod shaped gram negative bacteria when observed under microscope which are the feature of *Pseudomonas* genus.

Optimization studies:

Optimization studies were done to get a bacterial strain that is capable of degrading organophosphorus pesticide with the optimized parameters such as temperature, pH, substrate, time period as well as pesticide concentration which were published in (Gade Pavan et al., 2023). Available online at: <https://jazindia.com>

Pavan et al., 2023). Such potent colonies were grown in broth and incubated for further screening studies.

Screening studies of PDB:

Screening was done using sucrose as substrate in nutrient agar media and fresh colonies from the broth were spread and Chlorantraniliprole pesticide was also added of 10mg concentration and after incubating for 24 hours no zones were observed which indicated the organophosphate hydrolase enzyme activity of the bacterial isolates towards the pesticide degradation which were shown in figure 3.



Figure 3: No clear zones are seen due to pesticide degradation by bacterial hydrolase activity

Pesticide degrading bacteria are regarded as the better alternative for degrading pesticides which were accumulated in soil of agriculture fields. The bioremediation properties of microorganisms in particular bacteria is influenced by factors such as application of pesticide, their frequency of application, soil pH, stability of other flora, gap between the applications and half-life of the pesticide in the soil (Singh and Walker, 2006).

So to understand the metabolism of the PDBs for their activity, they need to be isolated and screened for their potency which was attempted in the present work through a method targeting 'oph' gene which produces organophosphate hydrolase enzyme which is produced when the bacteria degrades pesticides that belongs to the class organophosphorus which was evident by no zones and luxurious growth of bacteria even in the presence of pesticide as shown in figure 3, this luxurious growth is due to chemolithotrophic nutrition of PDBs.

It is reported that *Klebsiella aerogenes* degrade pesticides belonging to class methyl as well as dimethyl phosphates due to the presence of phosphomonoesterase and diesterase enzymes (Wolfenden and Spence, 1967). Even *Enterobacter* species were reported to use organophosphorus compounds and are able to metabolize them to simple compounds (Singh et al., 2003 and Singh et al., 2004). This OPH enzyme was first observed in *Falvobacterium* strain ATCC27551 as well as *Pseudomonas diminuta* and their coding genes on extrachromosomal plasmid were similar (Qionget al., 2010).

OPH enzyme isolated from *Pseudomonas* species have shown more thermal stability which helped the scientists to explore these organisms for understanding the enzyme activity toward pesticide degradation (Su et al., 2011). This very property has increased the interest in screening for the microbes related to *Pseudomonas* species which was shown in present work.

Conclusion:

Organophosphate hydrolase enzyme is the xenobiotic compound breaking down enzyme which specifically targets on organophosphorus compounds of pesticides such as Chlorantraniliprole which was used in the present study. This enzyme is naturally doing this activity in the vicinity of soil in agricultural fields where these class of pesticides are extensively used. Biological degradation of these compounds is necessary as they are bioaccumulating and biomagnifying themselves and taking much time to degrade in soil.

Pseudomonas species are one of the best organophosphorus compounds degraders as the OPH enzyme in them is naturally a thermostable one. This is evident that the obtaining of *Pseudomonas* in the present study as potent isolate for degradation of Chlorantraniliprole in optimized conditions.

Future studies are focused on testing the efficacy of OPH enzyme from *Pseudomonas* in degrading a wider range of class of pesticides by understanding the enzyme activity at molecular level.

References:

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