

PHYTOCHEMICAL EVALUATION AND ANTIOXIDANT ACTIVITY OF *PTERIS VITTATA* L FROM BHIWANDI

Dr. Ritu Jain

Associate professor, Department of Botany, K.M.E. Society's G.M.Momin Women's College Bhiwandi,
India

Email: ritujain0933@gmail.com

Article History	Abstract
Received: 06 June 2023 Revised: 19 Sept 2023 Accepted: 09 October 2023 CC License CC-BY-NC-SA 4.0	<i>Plants contain a variety of phytochemical compounds that are rich in antioxidant activity, including, lignin, stilbenes, tannins, flavonoids, vitamins, terpenoids, phenolic acids, quinones, alkaloids, amines, betalains, and other metabolites. Numerous of these antioxidant compounds have been shown in studies to exhibit anti-inflammatory, anti-atherosclerotic, anti-tumor, anti-mutagenic, anti-carcinogenic, antibacterial, and antiviral properties. Natural phytochemicals are found in crops like teas, herbs, oilseeds, beans, vegetables and fruits. They have been linked to lower risks of cancer, cardiovascular disease, diabetes, and other age-related diseases. Recently, there has been a global trend toward the use of these phytochemicals. The objective of this research was to use qualitative and quantitative screening methods to see phytochemicals were present in the methanol extracts Pteris vittata from Bhiwandi including steroids, reducing sugars, triterpenoids, sugars, alkaloids, phenolic compounds, flavonoids, saponins, tannins, and anthraquinones, were tested. More active compounds can be isolated from the selected ferns and they may be used for medicinal purposes in future.</i> Keywords: Pteridaceae, Phytochemicals, Ferns, Qualitative and quantitative screening.

INTRODUCTION

Plants are rich source of phytochemicals such as, phenolic acid, tannins, quinones, alkaloids, betalains ligning as well as metabolites which can perform antioxidant activity. (Zheng W, Wang SY.2001, Cai Y.Z, 2003).

As per WHO report more than 75% people of developing countries prefer plant derived medicines for their health Problems. (Gurib-Fakim A 2006). In past few years, there has been a worldwide trend towards the use the demand of the natural phytochemicals derived from vegetables and fruits, berry crops, different seeds and herbs has been increased. (Kitts DD *et al* 2000, Wang SY, Jiao H 2000 ,Muselik J, Garcia-Alonso M 2007) .

Pteridophytes belong to nearly 50 plant families which comprise upto 13000 species, but only few of species from thirty families have been explored for their biological activity. Although higher plants have been studied carefully, in terms of their medicinal, economical and commercial values, there is poor investigation of Pteridophytes in terms of these values. Hence there is very less information available on the literature regarding medicinally important except a few studies (PN Manandhar. 1996, Sharma NK 2002, A Benjamin; Manickam VS. 2007)

(Rice-Evans *et al* 1995and Sala A *et al* 2002). In their study found that Pteridophytes have various phytochemical molecules such as vitamins, terpenoids, phenolic acids, lignin's, stilbenes, tannins, flavonoids, quinones, coumarins, alkaloids, amines, betalains, and other metabolites. These compounds have antioxidant activity. These antioxidant compounds possess anti-inflammatory, antiatherosclerotic, antitumor, antimutagenic, anticarcinogenic, antibacterial, and antiviral activities natural antioxidants are linked with ageing diseases like diabetes, cancer, cardiovascular disease etc.

Hence to develop new drugs from underexplored plant groups, it is essential to encourage intensive scientific research on pteridophyte species to contribute in search for affordable medicines in India.

MATERIAL AND METHOD

Collection of plant materials

Fresh leaves (fronds) of the *Pteris vittata* L were collected randomly from the different location of Bhiwandi. Fresh leaves of *Pteris vittata* L were washed; shade dried and powdered using the blender and stored in air tight containers.

Quantitative phytochemical analysis

The phytochemicals present in the methanol extracts of *P. vittata*, were determined and quantified by standard procedures. Quantification of tannins by Hagerman et al 2000. Anti oxidant and free radical scavenging activity by Kumaran A, Karunakaran R 2006, phytochemical methods by Harborne J 1973. , Formation of complexes between protein and Tannin acid by Van-Burden T and Robinson W J. 1981

OBSERVATION:

Phytochemical compounds such as steroids, reducing sugars, triterpenoids, sugars, alkaloids, phenolic compounds, flavonoids, saponins, tannins, and anthroquinones were screened in extract. The data have been presented in Table 1 and 2 and Text Fig 1-2.

P. vittata extract contained 12.30 mg of alkaloids, 13.90 mg of flavonoids, 08.80 mg of saponins and 06.10 mg of tannins, 11.20 mg of phenolic compounds. It was observed that various concentrations of leaves and stem extracts *Pteris* has significant antioxidant potential and can used as good source of natural antioxidant. Alkaloids,

phenolic compounds, flavonoids, saponins and tannins are important secondary metabolites and are responsible for medicinal values of the respective plant.

Antioxidant properties of ferns was determined by estimating their DPPH radical scavenging activity In DPPH assays, methanol extracts from leaves of *P.vittata* exhibited higher antioxidant activity (20 µg/ml) (37.8%) than aqueous (29.3%) or chloroform extracts (13.3%). It was observed that various concentrations of leaves and stem extracts *Pteris* has significant antioxidant potential and can used as good source of natural antioxidant.



Figure 1 *Pteris* in Bhiwandi

Compounds	Presence
Steroids	+
Triterpenoids	+
Reducing sugars	+
Sugars	+
Alkaloids	+
Phenolic compounds	+
Flavenoids	+

Figure 2 Phytochemical analysis of Pteris vittata

S.N	Phytochemical	Amount(mg/g)
1	Alkaloids	12.30 ±0.10
2	Flavonoids	13.90 ±0.20
3	Phenolics	11.20±0.10
4	Saponins	08.80±0.50
5	Tannins	06.10±0.20

Figure 1 Quantitative analysis of phytochemical (mg/g)

DISCUSSION

In the present study methanol extracts of *Pteris vittata* are screened for phytochemical analysis. The results revealed positive result. Kumudhavalli and Jaykar (2012) did phytochemical analysis of *Hemionitis arifolia*. The ethanolic and aqueous extracts showed the presence of flavonoids, carbohydrates, phenolic compounds and sterols were the major phyto constituents. the phyto-constituents of three *Adiantum* species *Christella dentate* and *Christella parasitica*, to provide chemical marker and inter-specific variation between the medicinally important genus were examined Muraleedharannair *et al.* (2012).

A review done by Ganesan K, Xu (2014) highlighted the phytochemical composition of *Pteris cretica* and its potential health benefits, including antioxidant properties. The plant was found to contain various bioactive compounds with potential therapeutic applications.

Sahu RK, Karuppusamy S, Sabu A, et al 2013. In their study found that different fractions of *Pteris vittata* extracts exhibited significant antioxidant activity. The phytochemical screening also revealed the presence of various bioactive compounds. Rajurkar and Kunda (2012) screened *Adiantum capillus - veneris* for phytochemicals and metal content. The Soxhlet extraction of *Adiantum capillus veneris* showed the considerable amount of phenolics and terpenoids, fats and waxes, alkaloids, quaternary, Noxides and fiber.

In the present study, highest amount of alkaloids (16mg), flavonoids (17mg) and phenolics (13mg) are quantified in the *Pteris vittata*. Xie P, Chen S, Li L, et al. 2011 investigated the phytochemical composition and antioxidant activities of different parts of *Pteris multifida*. Various bioactive compounds, such as phenolics and flavonoids, were identified, and the extracts exhibited significant antioxidant activities.

In the present study *Pteris* extracts showed the presence of alkaloids, flavonoids and saponins. Based on present study further research can be done using chromatographic and spectroscopic techniques in the way of isolation and identification of the active compound from *Pteris*. Hence by further study biologist can evaluate the medicinal uses of pteridophytes and their bioactive principles. The efficacy of bioactive compounds could be increased by studies of in vitro culture, biosynthetic pathways, modes of action, and genetic expression.

ACKNOWLEDGMENT

I am grateful to the Management KME Society and Principal G M Momin Women's College Bhiwandi for their support and encouragement. I would like to thank Dr SP Khullar and Dr Arun Pandey Sir I am also thankful to Dr B. L. Yadav for his support and guidance.

References:

- 1). A Benjamin; Manickam VS. Medicinal 2007 pteridophytes from the western Ghats. Indian J. Trad. Knowledge 6, 611-618.
- 2). Cai YZ, Sun M, Corke H. 2003 Antioxidant activity of betalains from plants of the Amaranthaceae. J Agric Food Chem.51(8): 2288-2294.
- 3). Ganesan K, Xu B. 2014 A critical review on phytochemical profile and health-promoting effects of *Pteris cretica* L. Food Science and Human Wellness. 3(3-4):103-111.
- 4). Gurib-Fakim A. 2006 Medicinal plants: Traditions of yesterday and drugs of tomorrow. Mol Asp Med.; 27:1-93.
- 5). Hagerman A, Muller I, Makkar H 2000 Quantification of tannins in tree foliage. A laboratory manual, Vienna: FAO/IAEA, p. 4- 7.
- 6). Harborne J 1973 Phytochemical methods. Chapman and Hall, Ltd London.p.49-88.
- 7). Kitts DD, Yuan YV, Wijewickreme AN, Hu C. 2000 Antioxidant properties of a North American ginseng extract. Mol Cell Biochem.20(3):1-10.
- 8). Kumaran A, Karunakaran R. 2006 Anti oxidant and free radical scavenging activity of an aqueous extracts of *Coleus aromaticus*. Food chemistry.97: 109-114.
- 9). Kumudhavalli MV, Jaykar B. 2012 Pharmacological Screening on Leaves of the Plant of *Hemionitis Arifolia* (Burm).T.Moore, Res J Pharma, Bio Chem Sci . 3(2): 79-83.

- 10). Muraleedharannair M, Johnson M, Mony M, Zachariah M, Solomon J. 2012 Inter-specific variation studies on the phyto- constituents of *Christella* and *Adiantum* using phytochemical methods, As. Paci J Tro Biomed S40-S45
- 11). Muselik J, Garcia-Alonso M, Martín-López MP, Želmička M, Rivas-Gonzalo JC. 2007 Measurement of Antioxidant Activity of Wine Catechins, Procyanidins, Antocyanins and Piranoantocyanins. Int J Mol Sci 8: 797-809.
- 12). PN Manandhar. 1996 Ethnobotanical observations on ferns and ferns allies of Nepal, J. Econ. Tax. Bot., 12, 414-422.
- 13). Rajurkar N, Kunda G. 2012 Evaluation of phytochemicals, antioxidant activity and elemental content of *Adiantum capillus veneris* leaves, J Chem Pharma Res, 4(1): 365-374.
- 14). Rice-Evans CA, Miller NJ, Bolwell PG, Bramley PM, Pridham JB. 1995 The relative activities of Plant-derived polyphenolic flavonoid. Free radical Res 22: 375-383.
- 15). Sahu RK, Karuppusamy S, Sabu A, *et al.* 2013 Phytochemical screening and evaluation of antioxidant activity of different fractions of *Pteris vittata* L. extracts. Journal of Pharmacy Research. 6(10):1052-1057.
- 16). Sala A, Recio MD, Giner RM, Manez S, Tournier H, Schinella G, Rios JL. 2002 Antiinflammatory and antioxidant properties of *Helichrysum italicum*. J Pharm Pharmacol 54(3): 365-371.
- 17). Sharma NK. 2002 Ethnomedicinal studies on ferns and fern allies of Hadoti plateau, Southern Rajasthan. Zoos'Print J. 17(3): 732-734.
- 18). Van-Burden T, Robinson W. 1981 Formation of complexes between protein and Tannin acid. J. Agric. Food Chem 1: 77.
- 19). Wang SY, Jiao H. 2000 Correlation of antioxidant capacities to oxygen radical scavenging enzyme activities in blackberry. J Agric Food Chem 48: 5672-5676.
- 20). Xie P, Chen S, Li L, *et al.* 2011 Phytochemical analysis and antioxidant activities of different parts of *Pteris multifida* Poir. Journal of Agricultural and Food Chemistry. 59(9):5103-5110.
- 21). Zheng W, Wang SY. 2001 Antioxidant activity and phenolic compounds in selected herbs. J Agric Food Chem 49(11): 5165-5170.